## Current and future therapies for abnormal early embryogenesis with assisted reproductive technology

XIAOXIA WANG<sup>1,#</sup>; Zhongyuan YAO<sup>1,2,#</sup>; DI LIU<sup>1</sup>; Chunhong YU<sup>3,\*</sup>; Hui LI<sup>1,3,\*</sup>

<sup>1</sup> Department of Reproductive Medicine, Xiangya Hospital, Central South University, Changsha, 410000, China

<sup>2</sup> Center for Medical Genetics & Hunan Key Laboratory of Medical Genetics, School of Life Sciences, Central South University, Changsha, 410078, China

<sup>3</sup> Hunan Key Laboratory of Molecular Precision Medicine, Xiangya Hospital & Hunan Key Laboratory of Medical Genetics, School of Life Sciences, Central South University, Changsha, 410000, China

Key words: Oocyte maturation arrest, Preimplantation embryonic lethality, Gene mutation, Infertility, Embryo development

Abstract: Each stage of embryonic development, including normal gamete maturation, fertilization, zygotic genome activation, and cleavage, is crucial for human reproduction. Early embryo arrest is a common phenomenon. It is estimated that about 40%–70% of human embryos are arrested at early developmental stages. However, the exact mechanism remains largely uncertain. Embryos can be investigated *in vitro* by way of the development of *in vitro* fertilization/intracytoplasmic sperm injection. In addition to iatrogenic factors related to abnormal oocyte/embryo development, multiple gene mutations have been found to be involved in such phenotypes. Based on the knowledge of known etiological factors, several therapies are proposed to improve clinical outcomes. Here, we shed light on current and potential therapies for treating these conditions through reviewing articles and combining with our clinical and research experience.

According to reported data, infertility affects 15% of couples of reproductive age, and recurrent pregnancy loss affects approximately 5% of women of childbearing age globally (Sun et al., 2019; Zhou et al., 2021). Infertility and recurrent pregnancy loss are common disorders. However, their exact mechanisms remain unknown. The stages of embryonic development, including normal gamete maturation, fertilization, zygotic genome activation (ZGA), and cleavage, are of vital importance for uterine embryonic implantation and the development of a baby. It has been difficult to directly study human oocytes/embryos previously due to ethical challenges and the scarcity of research materials. However, with the help of in vitro fertilization/ intracytoplasmic sperm injection (IVF/ICSI), investigators are now able to observe developmentally abnormal human oocytes/embryos in vitro.

Oocyte/embryo developmental abnormality include internal (e.g., aneuploidy) and morphological factors that can be optimally addressed in the following stepwise fashion. Firstly, when it occurs in an IVF/ICSI cycle, a clinical

\*Address correspondence to: Hui Li, huili257@hotmail.com;

Chunhong Yu, yuchunhong@sklmg.edu.cn

Doi: 10.32604/biocell.2022.019731

doctor or embryologist must assess the patients' condition. Normal oocyte/embryonic development is impacted by body mass index (BMI), age, endometriosis, ovarian reserve, male sperm quality, and the couple's chromosomal makeup (Demko et al., 2016; Esteves et al., 2019; Lin et al., 2019; Le et al., 2021). Secondly, iatrogenic factors must be excluded. Noting the personalized treatment that doctors choose a controlled ovarian stimulation protocol according to clients is therefore important. It has been reported that a mild stimulation protocol such as progestin-primed ovarian stimulation (PPOS) improves the chances of attaining euploid embryos (La Marca et al., 2020). Furthermore, starting and maintenance dosage of gonadotropin (Gn) should be monitored and adjusted for each individual case. Sometimes, even the timing of human chorionic gonadotropin triggering influences oocyte maturation and embryo development. Thirdly, it is important to note that some adjuvant drugs, such as growth hormones, dehydroepiandrosterone, calcium, and vitamin D, have been implicated in improving oocyte and embryo quality (Nardo and Chouliaras, 2020). Finally, it is useful to consider that, in addition to these drugs, several modifications in in vitro maturation (IVM) media, such as melatonin supplementation, can improve levels of oocyte/ embryo maturation, although the effects are subtle (Nardo and Chouliaras, 2020; Li et al., 2021b).

www.techscience.com/journal/biocell



This work is licensed under a Creative Commons Attribution 4.0 International License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

<sup>&</sup>lt;sup>#</sup>The two authors contributed equally as first author

Received: 11 October 2021; Accepted: 21 December 2021

Notably, not all oocytes/embryos lead to live birth events. In fact, 40%–70% of human embryos are estimated to arrest at early developmental stages (Gardner and Lane, 1997). To complicate things even further, some infertile patients experience multiple IVF/ICSI cycles due to either the complete oocyte maturation arrest, preimplantation embryonic lethality, or recurrent implantation failures not attributable to any of the aforementioned factors. This leads to serious physiological and economic stress for patients.

A number of genes influencing early embryonic development have been identified in animal models, whereas the etiology in humans has remained largely elusive until recently. Due to advances in whole exome sequencing (WES), some abnormal oocyte/embryo development is considered as one of Mendelian disorders. Twenty-three genes have been confirmed to be related to abnormal human oocyte/embryo development. One gene links to oocyte death (PANX1 (Sang et al., 2019)). Another is tied to oocyte maturation arrest (TUBB8 (Feng et al., 2016), PALT2 (Chen et al., 2017b; Maddirevula et al., 2017a), TRIP3 (Zhang et al., 2020), CDC20 (Zhao et al., 2020) and MEI1 (Ben Khelifa et al., 2018)). Four links to genuine empty follicle syndrome (ZP1 (Zhou et al., 2019), ZP2 (Yang et al., 2021), ZP3 (Chen et al., 2017c) and LHCGR (Yariz et al., 2011)). Three are associated with an oocyte's lack of a zona pellucida (ZP1, ZP2 and ZP3 (Huang et al., 2014)). One gene has been implicated for thin zona pellucida (ZP2 (Dai et al., 2019)). Eight link to fertilization failure (ZP2 (Dai et al., 2019), PLCZ1 (Heytens et al., 2009), WEE2 (Sang et al., 2018), PALT2 (Wu et al., 2019), TUBB8 (Chen et al., 2017a), TLE6 (Alazami et al., 2015), CDC20 (Zhao et al., 2020) and NLRP5 (Li et al., 2021a)). Two have been implicated in zygotic cleavage failure (PADI6 (Maddirevula et al., 2017b) and BTG4 (Zheng et al., 2020)). Nine link to early embryo arrest (TLE6 (Alazami et al., 2015), NLRP2 (Mu et al., 2019), NLRP5 (Mu et al., 2019), PATL2 (Chen et al., 2017b), TUBB8 (Chen et al., 2017a), CDC20 (Zhao et al., 2020), KHDC3L (Wang et al., 2018), REC114 (Wang et al., 2020), MEI1 (Dong et al., 2021) and MOS (Zhang et al., 2021b)). One has been associated with pronuclear fusion failure (CHK1 (Zhang et al., 2021a)). Five have been tied to recurrent hydatidiform moles (KDHC3L (Wang et al., 2018), PADI6 (Maddirevula et al., 2017b), NLRP5 (Rezaei et al., 2021), NLRP7 (Rezaei et al., 2021) and MEI1 (Nguyen et al., 2018)), and so on. As a result of these new insights, an expanding spectrum of mutation sites and phenotypes has been identified. It seems that different phenotypes of infertility can be linked to one gene, and one causative gene may induce several phenotypes. This indicates that the pathways of oocyte/embryo development are very interconnected and complex. Reproductive doctors and genetic counselors must be aware of the links among infertility, miscarriages, and aberrant chromosomal makeup. For patients with multiple unexplained IVF/ICSI failures, WES and Sanger sequencing combined with family history must be ascertained to confirm whether they carry any of the reported pathogenic genes. In cases where pathogenic variants are identified, doctors can offer genetic and reproductive counselling options to affected couples. Sperminduced oocyte activation failure has been successfully

treated by way of artificial oocyte activation (AOA) with no increase in birth defect rate (Long *et al.*, 2020). However, for other conditions, oocyte/sperm donation should be suggested because patients carrying these mutations rarely give birth, yet it might still be a feasible option to enable carriers to have their own children (Sang *et al.*, 2021). Moreover, it should be mentioned that abnormal oocyte/ embryo development cannot be completely explained by the aforementioned genes. Thus, WES for these patients can help identify novel pathological genes, which would thus pave the way for future diagnosis and treatment.

Gene therapy may soon become a promising treatment option. In 2020, Zhao *et al.* (2020) injected CDC20 complementary RNAs (cRNAs) into oocytes in maturation arrest (Zhao *et al.*, 2020). All injected oocytes were successfully fertilized, and developed into embryos, in which case the preimplantation genetic screening showed that one of the blastocytes was normal (Zhao *et al.*, 2020). In addition, injections of human *PLCZ1* cRNAs or recombinant protein were able to trigger Ca<sup>2+</sup> oscillation, which aided oocytes' development to the blastocyte stage (Rogers *et al.*, 2004; Yoon *et al.*, 2012). Exogenous cRNAs and protein complements can help early embryonic transition to the next development phase. However, it remains a big question whether such cRNAs or protein injections will ultimately adversely impact progeny, despite their rapid rate of metabolization.

For embryos experiencing causally unknown recurrent implantation failure, preimplantation genetic testinganeuploidy (PGT-A) will soon be applicable for screening euploid embryos to increase their chances of implantation. PGT is an invasive technology that requires the suction of 5–8 trophectoderm (TE) cells for effective detection. Furthermore, it does not lend to a direct means of assessing the inner cell mass (ICM). Recently, noninvasive PGT-A (niPGT-A), a new technology, has been proposed for analyzing cell-free DNA in the spent culture media of human blastocytes to directly assess for ICM (Huang et al., 2019). However, it includes expelled abnormal chromosomes from TE/ICM, which reduces the reliability of the technology. Fluorescence lifetime imaging microscopy (FLIM) is another technology used to measure metabolic characteristics of embryos to identify the ploidy of an embryo (Sanchez et al., 2019). Nonetheless, presently the researches is still solely focused on mouse oocytes/embryos and is yet to be applied to human embryos.

In conclusion, oocyte/embryonic development is influenced by both the personal chromosomal status of patients and iatrogenic factors. To date, a series of genetic contributors has been identified by the way of combing clinical phenotypes and WES. These genes can be potential biomarkers for diagnosing patients with embryo developmental abnormalities, and may thus be a viable direction for future therapy. Although NiPGT-A and FLIM are noninvasive technologies for identifying euploidies, they have their disadvantages. We anticipate that new noninvasive technologies will be developed to replace PGT-A for direct assessment of ICM. By these means, future reproductive physicians will not be confined to simply optimizing clinical treatments so as to avoid iatrogenic factors. Rather they will also be equipped to diagnose such patients and thereby point them towards a new range of helpful avenues for assisting them.

Acknowledgement: We gratefully acknowledge Prof. Kai Yuan and the members of the Yuan Lab for their helpful discussions.

Author Contribution: The authors confirm contribution to the paper as follows: study conception and design: Hui Li and Chunhong Yu; data collection: Di Liu; draft manuscript preparation: Xiaoxia Wang and Zhongyuan Yao. All authors reviewed the results and approved the final version of the manuscript.

Ethics Approval: Not applicable.

**Funding Statement:** This project was supported by The Third Batch of China Post-Doctoral Special Funding (Grant 2021TQ0372 to H.L.), and the Key Research and Development Plan of Hunan Province (Grant 2021SK2028 to H.L.).

**Conflicts of Interest:** The authors declare that they have no conflicts of interest to report regarding the present study.

## References

- Alazami AM, Awad SM, Coskun S, Al-Hassan S, Hijazi H et al. (2015). *TLE6* mutation causes the earliest known human embryonic lethality. *Genome Biology* 16: 108. DOI 10.1186/ s13059-015-0792-0.
- Ben Khelifa M, Ghieh F, Boudjenah R, Hue C, Fauvert D et al. (2018). A MEI1 homozygous missense mutation associated with meiotic arrest in a consanguineous family. Human Reproduction 33: 1034–1037. DOI 10.1093/humrep/dey073.
- Chen B, Li B, Li D, Yan Z, Mao X et al. (2017a). Novel mutations and structural deletions in *TUBB8*: Expanding mutational and phenotypic spectrum of patients with arrest in oocyte maturation, fertilization or early embryonic development. *Human Reproduction* **32**: 457–464. DOI 10.1093/humrep/dew322.
- Chen B, Zhang Z, Sun X, Kuang Y, Mao X et al. (2017b). Biallelic mutations in *PATL2* cause female infertility characterized by oocyte maturation arrest. *American Journal of Human Genetics* **101**: 609–615. DOI 10.1016/j.ajhg.2017.08.018.
- Chen T, Bian Y, Liu X, Zhao S, Wu K et al. (2017c). A recurrent missense mutation in *ZP3* causes empty follicle syndrome and female infertility. *American Journal of Human Genetics* **101**: 459–465. DOI 10.1016/j.ajhg.2017.08.001.
- Dai C, Hu L, Gong F, Tan Y, Cai S et al. (2019). ZP2 pathogenic variants cause in vitro fertilization failure and female infertility. Genetics in Medicine 21: 431–440. DOI 10.1038/ s41436-018-0064-y.
- Demko ZP, Simon AL, McCoy RC, Petrov DA, Rabinowitz M (2016). Effects of maternal age on euploidy rates in a large cohort of embryos analyzed with 24-chromosome single-nucleotide polymorphism-based preimplantation genetic screening. *Fertility and Sterility* 105: 1307–1313. DOI 10.1016/j. fertnstert.2016.01.025.
- Dong J, Zhang H, Mao X, Zhu J, Li D et al. (2021). Novel biallelic mutations in *MEI1*: Expanding the phenotypic spectrum to human embryonic arrest and recurrent implantation failure. *Human Reproduction* **36**: 2371–2381.
- Esteves SC, Carvalho JF, Bento FC, Santos J (2019). A novel predictive model to estimate the number of mature oocytes required for obtaining at least one euploid blastocyst for transfer in couples undergoing *in vitro* fertilization/ intracytoplasmic sperm injection: The ART calculator.

*Frontiers in Endocrinology* **10**: 168. DOI 10.3389/ fendo.2019.00099.

- Feng R, Sang Q, Kuang Y, Sun X, Yan Z et al. (2016). Mutations in TUBB8 and human oocyte meiotic arrest. New England Journal of Medicine 374: 223–232. DOI 10.1056/NEJMoa1510791.
- Gardner DK, Lane M (1997). Culture and selection of viable blastocysts: A feasible proposition for human IVF? *Human Reproduction Update* **3**: 367–382. DOI 10.1093/humupd/3.4.367.
- Heytens E, Parrington J, Coward K, Young C, Lambrecht S et al. (2009).
  Reduced amounts and abnormal forms of phospholipase C zeta (PLC) in spermatozoa from infertile men. *Human Reproduction* 24: 2417–2428. DOI 10.1093/humrep/dep207.
- Huang H, Lv C, Zhao Y, Li W, He X et al. (2014). Mutant ZP1 in familial infertility. *New England Journal of Medicine* **370**: 1220–1226. DOI 10.1056/NEJMoa1308851.
- Huang L, Bogale B, Tang Y, Lu S, Xie XS, Racowsky C (2019). Noninvasive preimplantation genetic testing for aneuploidy in spent medium may be more reliable than trophectoderm biopsy. Proceedings of the National Academy of Sciences of the United States of America 116: 14105–14112. DOI 10.1073/pnas.1907472116.
- La Marca A, Capuzzo M, Sacchi S, Imbrogno MG, Spinella F et al. (2020). Comparison of euploidy rates of blastocysts in women treated with progestins or GnRH antagonist to prevent the luteinizing hormone surge during ovarian stimulation. *Human Reproduction* **35**: 1325–1331. DOI 10.1093/humrep/deaa068.
- Le MT, Nguyen TV, Nguyen TTT, Nguyen HTT, Le DD, Nguyen VQH (2021). Predictive significance of sperm DNA fragmentation testing in early pregnancy loss in infertile couples undergoing intracytoplasmic sperm injection. *Research and Reports in Urology* 13: 313–323. DOI 10.2147/RRU.S315300.
- Li M, Jia M, Zhao X, Shi R, Xue X (2021a). A new NLRP5 mutation causes female infertility and total fertilization failure. *Gynecological Endocrinology* 37: 283–284. DOI 10.1080/ 09513590.2020.1832069.
- Li X, Mu Y, Elshewy N, Ding D, Zou H et al. (2021b). Comparison of IVF and IVM outcomes in the same patient treated with a modified IVM protocol along with an oocytes-maturing system containing melatonin: A pilot study. *Life Sciences* 264: 118706. DOI 10.1016/j.lfs.2020.118706.
- Lin Y, Yang P, Chen Y, Zhu J, Zhang X et al. (2019). Factors inducing decreased oocyte maturation rate: A retrospective analysis of 20,939 ICSI cycles. Archives of Gynecology and Obstetrics 299: 559–564. DOI 10.1007/s00404-018-4958-3.
- Long R, Wang M, Yang QY, Hu SQ, Zhu LX et al. (2020). Risk of birth defects in children conceived by artificial oocyte activation and intracytoplasmic sperm injection: A metaanalysis. *Reproductive Biology and Endocrinology* 18: 17. DOI 10.1186/s12958-020-00680-2.
- Maddirevula S, Coskun S, Alhassan S, Elnour A, Alsaif HS et al. (2017a). Female infertility caused by mutations in the oocyte-specific translational repressor *PATL2*. *American Journal of Human Genetics* **101**: 603–608. DOI 10.1016/j. ajhg.2017.08.009.
- Maddirevula S, Coskun S, Awartani K, Alsaif H, Abdulwahab FM et al. (2017b). The human knockout phenotype of *PADI6* is female sterility caused by cleavage failure of their fertilized eggs. *Clinical Genetics* **91**: 344–345. DOI 10.1111/cge.12866.
- Mu J, Wang W, Chen B, Wu L, Li B et al. (2019). Mutations in *NLRP2* and *NLRP5* cause female infertility characterised by early

embryonic arrest. *Journal of Medical Genetics* **56**: 471–480. DOI 10.1136/jmedgenet-2018-105936.

- Nardo L, Chouliaras S (2020). Adjuvants in IVF-evidence for what works and what does not work. Upsala Journal of Medical Sciences 125: 144–151. DOI 10.1080/03009734.2020.1751751.
- Nguyen NMP, Ge Z, Reddy R, Fahiminiya S, Sauthier P et al. (2018). Causative mutations and mechanism of androgenetic hydatidiform moles. *American Journal of Human Genetics* **103**: 740–751. DOI 10.1016/j.ajhg.2018.10.007.
- Rezaei M, Suresh B, Bereke E, Hadipour Z, Aguinaga M et al. (2021). Novel pathogenic variants in *NLRP7*, *NLRP5*, and *PADI6* in patients with recurrent hydatidiform moles and reproductive failure. *Clinical Genetics* **99**: 823–828. DOI 10.1111/cge.13941.
- Rogers NT, Hobson E, Pickering S, Lai FA, Braude P et al. (2004). Phospholipase  $C\zeta$  causes  $Ca^{2+}$  oscillations and parthenogenetic activation of human oocytes. *Reproduction* **128**: 697–702. DOI 10.1530/rep.1.00484.
- Sanchez T, Venturas M, Aghvami SA, Yang X, Fraden S et al. (2019). Combined noninvasive metabolic and spindle imaging as potential tools for embryo and oocyte assessment. *Human Reproduction* 34: 2349–2361. DOI 10.1093/humrep/dez210.
- Sang Q, Li B, Kuang Y, Wang X, Zhang Z et al. (2018). Homozygous mutations in WEE2 cause fertilization failure and female infertility. American Journal of Human Genetics 102: 649– 657. DOI 10.1016/j.ajhg.2018.02.015.
- Sang Q, Zhang Z, Shi J, Sun X, Li B et al. (2019). A pannexin 1 channelopathy causes human oocyte death. Science Translational Medicine 11: eaav8731. DOI 10.1126/ scitranslmed.aav8731.
- Sang Q, Zhou Z, Mu J, Wang L (2021). Genetic factors as potential molecular markers of human oocyte and embryo quality. *Journal of Assisted Reproduction and Genetics* 38: 993– 1002. DOI 10.1007/s10815-021-02196-z.
- Sun H, Gong TT, Jiang YT, Zhang S, Zhao YH et al. (2019). Global, regional, and national prevalence and disability-adjusted lifeyears for infertility in 195 countries and territories, 1990– 2017: Results from a global burden of disease study, 2017. *Aging (Albany NY)* **11**: 10952–10991. DOI 10.18632/ aging.102497.
- Wang W, Dong J, Chen B, Du J, Kuang Y et al. (2020). Homozygous mutations in REC114 cause female infertility characterised by multiple pronuclei formation and early embryonic arrest. *Journal of Medical Genetics* 57: 187–194. DOI 10.1136/ jmedgenet-2019-106379.
- Wang X, Song D, Mykytenko D, Kuang Y, Lv Q et al. (2018). Novel mutations in genes encoding subcortical maternal complex proteins may cause human embryonic developmental arrest. *Reproductive BioMedicine Online* 36: 698–704. DOI 10.1016/j.rbmo.2018.03.009.

- Wu L, Chen H, Li D, Song D, Chen B et al. (2019). Novel mutations in *PATL2*: expanding the mutational spectrum and corresponding phenotypic variability associated with female infertility. *Journal of Human Genetics* 64: 379–385. DOI 10.1038/s10038-019-0568-6.
- Yang P, Chen T, Liu Y, Hou Z, Wu K et al. (2021). The critical role of ZP genes in female infertility characterized by empty follicle syndrome and oocyte degeneration. Fertility and Sterility 115: 1259–1269. DOI 10.1016/j.fertnstert.2020.11.003.
- Yariz KO, Walsh T, Uzak A, Spiliopoulos M, Duman D et al. (2011). Inherited mutation of the luteinizing hormone/ choriogonadotropin receptor (LHCGR) in empty follicle syndrome. *Fertility and Sterility* **96**: e125–e130. DOI 10.1016/j.fertnstert.2011.05.057.
- Yoon SY, Eum JH, Lee JE, Lee HC, Kim YS et al. (2012). Recombinant human phospholipase C zeta 1 induces intracellular calcium oscillations and oocyte activation in mouse and human oocytes. *Human Reproduction* 27: 1768– 1780. DOI 10.1093/humrep/des092.
- Zhang H, Chen T, Wu K, Hou Z, Zhao S et al. (2021a). Dominant mutations in *CHK1* cause pronuclear fusion failure and zygote arrest that can be rescued by CHK1 inhibitor. *Cell Research* 31: 814–817. DOI 10.1038/s41422-021-00507-8.
- Zhang YL, Zheng W, Ren P, Hu H, Tong X et al. (2021b). Biallelic mutations in MOS cause female infertility characterized by human early embryonic arrest and fragmentation. *EMBO Molecular Medicine* 13: e14887. DOI 10.15252/ emmm.202114887.
- Zhang Z, Li B, Fu J, Li R, Diao F et al. (2020). Bi-allelic missense pathogenic variants in *TRIP13* cause female infertility characterized by oocyte maturation arrest. *American Journal of Human Genetics* 107: 15–23. DOI 10.1016/j.ajhg.2020.05.001.
- Zhao L, Xue S, Yao Z, Shi J, Chen B et al. (2020). Biallelic mutations in *CDC20* cause female infertility characterized by abnormalities in oocyte maturation and early embryonic development. *Protein & Cell* 11: 921–927. DOI 10.1007/ s13238-020-00756-0.
- Zheng W, Zhou Z, Sha Q, Niu X, Sun X et al. (2020). Homozygous mutations in *BTG4* cause zygotic cleavage failure and female infertility. *American Journal of Human Genetics* 107: 24–33. DOI 10.1016/j.ajhg.2020.05.010.
- Zhou Q, Xiong Y, Qu B, Bao A, Zhang Y (2021). DNA methylation and recurrent pregnancy loss: A mysterious compass? *Frontiers in Immunology* 12: 738962. DOI 10.3389/ fimmu.2021.738962.
- Zhou Z, Ni C, Wu L, Chen B, Xu Y et al. (2019). Novel mutations in ZP1, ZP2, and ZP3 cause female infertility due to abnormal zona pellucida formation. Human Genetics 138: 327–337. DOI 10.1007/s00439-019-01990-1.