

## Biomechanical Characteristics Closely Related with Immune Functions of Dendritic Cells

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**Abstract:** As potent antigen presenting cells, dendritic cells (DCs) are utilized to deliver the signals essential for the initiation of immune responses. The motility of DCs is crucial for migration of immature DCs (imDCs) in peripheral tissue and the interaction between mature DCs (mDCs) and naïve T cells in the secondary lymph node. From biomechanical viewpoint, the deformability of cells is necessary for their motility. Deformation of cells can be divided into active deformation (e.g. chemotaxis) and passive deformation (e.g. migration under shear stress of blood flow). However, there is no detailed study on the deformability of DCs including imDCs and mDCs. In this study, the active and passive deformations of DCs under various culture conditions were respectively measured by atomic force microscopy and micropipette. The results showed that DCs at different differentiation stages displayed various deformabilities, their passive and active deformation depended on matrix stiffness, mDCs had higher active deformation and Young's modulus as well as lower passive deformation. It's significant for further understanding the immunoregulatory functions of DCs.

Keywords: Dendritic cells; biomechanical characteristics; deformability; immunoregulatory function

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## References

- 1. Chien S. Red cell deformability and its relevance to blood flow. *Annual Review of Physiology* **1987**, 49: 177-192.
- Xiao L, Liu Y, Chien S, Fu B. Simulation of Deformation and Aggregation of Two Red Blood Cells in a Stenosed Microvessel by Dissipative Particle Dynamics. *Cell Biochemistry and Biophysics* 2016, 74(4): 513-525.
- 3. Maciaszek JL, Lykotrafitis G. Sickle cell trait human erythrocytes are significantly stiffer than normal. *Journal of Biomechanics* **2011**, 44(4): 657-661.
- 4. Dulinska I, Targosz M, Strojny W, Lekka M, Czuba P et al. Stiffness of normal and pathological erythrocytes studied by means of atomic force microscopy. *Journal of Biochemical and Biophysical Methods* **2006**, 66: 1-11.
- 5. Kohn JC, Lampi MC, Reinhart-King CA. Age-related vascular stiffening: causes and consequences. *Frontiers in genetics* **2015**, 6: 112.
- 6. Koch CG, Duncan AI, Figueroa P, Dai L, Sessler DI et al. Real Age: Red Blood Cell Aging During Storage. *Annals of Thoracic Surgery* **2019**, 107(3): 973-980.
- 7. Musielak M. Red blood cell-deformability measurement: review of techniques. *Clinical Hemorheology and Microcirculation* **2009**, 42(1): 47-64.
- 8. Barns S, Balanant MA, Sauret E, Flower R, Saha S et al. Investigation of red blood cell mechanical properties using AFM indentation and coarse-grained particle method. *BioMedical Engineering OnLine* **2017**, 16(1): 140.
- 9. Ciasca G, Papi M, Di Claudio S, Chiarpotto M, Palmieri V et al. Mapping viscoelastic properties of healthy and pathological red blood cells at the nanoscale level. *Nanoscale* **2015**, 7(40): 17030-17037.

- 10. Arashiki N, Kimata N, Manno S, Mohandas N, Takakuwa Y. Membrane peroxidation and methemoglobin formation are both necessary for band 3 clustering: mechanistic insights into human erythrocyte senescence. *Biochemistry* **2013**, 52(34): 5760-5769.
- 11. Bosman GJ, Willekens FL, Werre JM. Erythrocyte aging: a more than superficial resemblance to apoptosis? *Cell Biochemistry and Biophysics* **2005**, 16: 1-8.
- 12. Andrews DA, Yang L, Low PS. Phorbol ester stimulates a protein kinase C-mediated agatoxin-TK-sensitive calcium permeability pathway in human red blood cells. *Blood* **2002**, 100(9): 3392-3399.