

Multi-strain Inoculation with PGPR Producing ACC Deaminase is More Effective Than Single-strain Inoculation to Improve Wheat (*Triticum aestivum*) Growth and Yield

Muhammad Zafar-ul-Hye^{1,*}, Misbah Batool Zahra¹, Subhan Danish¹, Mazhar Abbas²,
Abdur Rehman¹, Muhammad Naeem Akbar¹, Ayesha Iftikhar¹, Mehreen Gul¹, Ifat Nazir¹,
Maria Abid¹, Muhammad Tahzeeb-ul-Hassan¹ and Maria Murtaza³

¹Department of Soil Science, Faculty of Agricultural Sciences and Technology, Bahauddin Zakariya University, Multan, 60800, Pakistan

²Institute of Horticultural Sciences, University of Agriculture, Faisalabad, 38040, Pakistan

³Institute of Soil and Environmental Sciences, University of Agriculture, Faisalabad, 38040, Pakistan

*Corresponding Author: Muhammad Zafar-ul-Hye. Email: zafarulhyegondal@yahoo.com

Received: 24 October 2019; Accepted: 18 December 2019

Abstract: Rhizosphere bacteria that colonize plant roots and confer beneficial effects are referred as plant growth promoting rhizobacteria (PGPR). Among all PGPR, some rhizobacteria have an ability to produce ACC deaminase enzyme. This enzyme catalyzes stress ACC into a-ketobutyrate and ammonia instead of letting it to be converted to ethylene. Ethylene level rises in plants under stress conditions i.e., drought, salinity, poor soil fertility etc. As poor soil fertility is a big hurdle to achieve the optimum yield of crops, inoculation of ACC deaminase PGPR can overcome this problem to some extent. The aim of the current study was to examine the influence of multi-strain and single-strain inoculation of different ACC deaminase producing PGPR on wheat growth and yield. There were three PGPR strains, *Enterobacter cloacae*, *Serratia ficaria* and *Burkholderia phytofirmans* which were used as consortia and single-strain inoculations. The results showed that inoculation of *E. cloacae* + *S. ficaria* + *B. phytofirmans* significantly increased plant height (63%), spike length (61%), number of spikelets spike⁻¹ (61%), number of grains spike⁻¹ (131%), 1000 grains weight (33%), grains yield (71%), straw yield (71%) and biological yield (68%) of wheat as compared to control. A significant improvement in N (37 and 200%), P (46 and 166%) and K (39 and 61%) of seeds and shoot respectively, validated the efficacious and more effective role of multi-strain (*E. cloacae* + *S. ficaria* + *B. phytofirmans*) inoculation over control. It is obviously concluded that multi-strain ACC deaminase producing PGPR inoculation is a better approach as compared to single-strain inoculation for the improvement in growth and yield of wheat.

Keywords: Ethylene; growth attributes; nutrients; consortium; rhizobacteria



This work is licensed under a Creative Commons Attribution 4.0 International License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

1 Introduction

Plant growth is directly linked with nutrients and water holding potential of soil [1]. High volatilization of ammonia, leaching of nitrate [2] and immobilization of phosphorus and potassium decrease the fertility level of soils [3]. Poor soil fertility status is a main hurdle to achieve the target of getting maximum yield from crops. Furthermore, intensive cultivation of exhaustive crops has depleted the nutrient concentrations in the soils [4]. Low availability of nutrients is associated with poor growth, development of plants [5–6].

Ethylene (C₂H₄) is a plant-signaling molecule which involves in the germination of seeds, root development and elongation, flower senescence, leaf abscission and fruit ripening. It is produced in a two-step process i.e., (1) enzymatic conversion of S-adenosyl methionine (SAM) to 1-amino cyclopropane-1-carboxylic acid (ACC); (2) conversion of ACC to ethylene, which is catalyzed by ACC-oxidase [7]. However, synthesis of endogenous ethylene level is significantly enhanced upon exposure of plants to abiotic stresses including low soil fertility [8–9].

Currently, various strategies are in use to maximize the yield of crops under nutritional stress using chemical fertilizers, green manures, farmyard manures and bio-fertilizers etc. In this context, the combined use of bio-fertilizers and inorganic fertilizers is more effective and environmental friendly approach [10–11]. It has been observed that rhizosphere is a zone of intense microbial activity immediately adjacent to the plant root and nutrient rich environment due to release of sugars, amino acids, organic acids, plant hormones and enzymes [12]. The plant growth promoting rhizobacteria (PGPR) can secrete multiple metabolites and enzymes that confer beneficial effects on plants growth [13–14].

In addition, the PGPR containing ACC deaminase have been found to be effective enough in enhancing the resistance against abiotic stresses and to promote the health of plants [15–16]. ACC deaminase is a polymeric enzyme which is dependent on pyridoxal 5-phosphate (PLP) [17]. It is established that the PGPR containing ACC deaminase can reduce the accumulation of ethylene by breaking cyclo-propanoid amino acid ACC (ethylene precursor), into intermediate compounds, ammonia and α -ketobutyrate [8]. The ACC deaminase producing PGPR enhance the uptake of N, P and K too especially in cereal crops like wheat and maize [18–19].

Across the globe, wheat is consumed as a staple food. It is widely cultivated due to its high nutritional value and capability to grow under a vast range of climatic conditions. Wheat shares 44% of daily human intake of iron (Fe) and 25% of zinc (Zn) [20]. In Pakistan, wheat is cultivated over an area of 8,740 thousand hectares with the production of 25,195 million tonnes. However, the yield of wheat is yet below the demand of exponentially increasing population [21]. That's why current experiment was conducted with the aim to examine the influence of single and multi-strain inoculation of ACC deaminase producing PGPR along with chemical fertilizers on growth and yield of wheat. It is hypothesized that multi-strain inoculation of ACC-deaminase producing PGPR would be a better approach to improve the growth and yield of wheat comparative to single-strain inoculation.

2 Materials and Methodology

2.1 Experimental Site

A field study was conducted in the experimental area of the Department of the Soil Science, Bahauddin Zakariya University Multan, Pakistan. For determination of soil texture, hydrometer method was used [22]. The pH and EC of soil were noted by making soil paste with 1:1 and 1:10 soil-water ratio respectively on pH and EC meters accordingly. The available P was found out through the protocol described by Olsen's [23], Extractable K was determined by following Nadeem et al. [24] and Walkley [25] protocol was followed for soil organic matter determination. The pre-experimental soil characteristics are provided in [Tab. 1](#).

Table 1: Physio-chemical characterization of soil

| Parameters | Unit | Values |
|--------------------------|---------------------|--------|
| Physical Analysis | | |
| Sand | % | 35 |
| Silt | % | 35 |
| Clay | % | 30 |
| Saturation percentage | % | 47 |
| Textural class | Clay loam | |
| Chemical Analysis | | |
| pHs | | 8.60 |
| ECe | dS m ⁻¹ | 2.30 |
| TDS | mg kg ⁻¹ | 1.15 |
| OM | % | 0.40 |
| Total nitrogen | % | 0.05 |
| Available phosphorus | mg kg ⁻¹ | 5.6 |
| Extractable potassium | mg kg ⁻¹ | 130 |

2.2 Treatments

There were eight treatments with three replications. The treatments included (1) un-inoculated control, (2) *Enterobacter cloacae* (3) *Serratia ficaria*, (4) *Burkholderia phytofirmans*, (5) *Enterobacter cloacae* + *Serratia ficaria*, (6) *Enterobacter cloacae* + *Burkholderia phytofirmans*, (7) *Serratia ficaria* + *Burkholderia phytofirmans* and (8) *Enterobacter cloacae* + *Serratia ficaria* + *Burkholderia phytofirmans*. The NPK fertilizers as 120, 100 and 60 kg ha⁻¹ respectively, were applied along with every treatment including control.

2.3 Bacterial Inoculation

Peat was used as a standard carrier for bacterial inoculation. Peat was collected from a local nursery and sterilized at 121°C for 21 min. For seed inoculation, the DF media was prepared without agar [26]. Uniform cell density, 10⁸-10⁹ CFU mL⁻¹ was achieved prior to seed inoculation in case of every single and multi-strain inoculation.

2.4 PGPR Strains

The PGPR strains, *Enterobacter cloacae* (W6), *Serratia ficaria* (W10) and *Burkholderia phytofirmans* (PsJN) were obtained from Laboratory of Soil Microbiology and Biochemistry, Institute of Soil and Environmental Sciences, University of Agriculture Faisalabad, Pakistan. All the strains were positive in ACC deaminase activity.

2.5 Wheat Variety and Sowing

Wheat cultivar used in the trial was AAS-2008 (a developed variety of Wheat for lower Punjab). Sowing was done by using dibbler (recommended 9 inch row to row distance) for sowing of seeds of Wheat. Each plot size was 12 m² (4 m × 3 m). Recommended row to row (R × R) distance i.e., 9 inch was maintained.

2.6 Irrigation and Fertilizer Application

Recommended number of irrigation (4-5) was applied throughout the research period in this field experiment, as per requirement. The NPK Fertilizers as urea, DAP and SOP were added at the rate of 120, 60 and 60 kg ha⁻¹ as recommended fertilizer doses [40].

2.7 Harvesting

Plants were harvested at the time of maturity. Plant height was measured at physiological maturity stage. Five tillers were chosen at random from each replication. Length of five random selected spikes was measured from each replication. Number of spikelet spike⁻¹ was also counted from the five randomly selected spike. Randomly harvested tillers were threshed separately. After separating grains from straw, number of grains per spike was counted. Five samples having 100 grains were taken from each plot and weighed. Then the weight of 100 grains was converted into 1000-grain weight by multiplication. Five plants were selected on random basis and the weight of whole produce of a plant was noted. Biological yield was calculated and adjusted to kg ha⁻¹ unit by multiplying the one meter square biological yield with 10,000 value. With the help of one meter square (1 m²) quadrat/frame, plants were harvested and sun dried for three days. Then they were threshed manually and after separating the grains and straws, grains were weighed and adjusted to kg ha⁻¹ unit by multiplying the one meter square grain yield by 10,000 value.

$$1 \text{ m}^2 \times 10,000 = \text{kg ha}^{-1}$$

Straw yield was calculated as:

$$\text{Straw yield} = \text{Biological yield} - \text{Grain yield}$$

2.8 Nutrients Analyses

For determination of nitrogen, potassium and phosphorus contents in grain and straw samples wet digestion was done as described by Wolf [27]. The potash was determined with flamephotometer (Jenway PFP-7), using standard curve [24]. Phosphorus was determined by spectrophotometer using standard curve [28]. Nitrogen was determined by Kjeldahl's method described by Jackson [29].

2.9 Statistical Analyses

Collected data from treatments on various parameters were arranged and analyzed by ANOVA (analysis of variance) and their means were compared with Tukey's test at 0.05% probability [30].

3 Results

3.1 Growth Attributes

Results revealed that single and multi-strains inoculation significantly affected wheat plant height, spike length, number of spikelets spike⁻¹ and number of grains spike⁻¹. Inoculation of *E. cloacae* × *S. ficaria* × *B. phytofirmans* remained significantly better over control for plant height, spike length, number of spikelets spike⁻¹ and number of grains spike⁻¹. No significant change was observed among each other where *E. cloacae* + *S. ficaria* and *S. ficaria* + *B. phytofirmans* were used for plant height and spike length but *E. cloacae* + *B. phytofirmans* performed significantly better over *S. ficaria* + *B. phytofirmans* for number of spikelets spike⁻¹ and number of grains spike⁻¹ (Tab. 2). It was also observed that sole *E. cloacae*, *S. ficaria* and *B. phytofirmans* differed significantly over control for plant height, spike length, number of spikelets spike⁻¹ and number of grains spike⁻¹. Among sole inoculation, *B. phytofirmans* remained significantly better from *E. cloacae* and *S. ficaria* for number of spikelets spike⁻¹ and number of grains spike⁻¹. Maximum increases of 62, 61, 61 and 131 % were noted in plant height, spike length, number of spikelets spike⁻¹ and number of grains spike⁻¹ respectively over control where *E. cloacae* + *S. ficaria* + *B. phytofirmans* was applied.

Table 2: Effect of single and multistrains inoculation of ACC deaminase producing PGPR on growth attributes of wheat

| Treatments | Plant Height (cm) | Spike Length (cm) | Number of spikelets spike ⁻¹ | Number of grains spike ⁻¹ |
|--|-------------------|-------------------|---|--------------------------------------|
| Control | 55.00 E | 8.67 E | 14.67 F | 25.67 H |
| <i>E. cloacae</i> | 66.33 CD | 11.00 CD | 18.00 E | 46.00 F |
| <i>S. ficaria</i> | 62.00 DE | 9.33 DE | 19.33 D | 42.67 G |
| <i>B. phytofirmans</i> | 66.67 CD | 11.33 BCD | 20.00 C | 48.33 E |
| <i>E. cloacae</i> + <i>S. ficaria</i> | 73.00 BC | 11.67 BC | 20.00 C | 50.00 C |
| <i>E. cloacae</i> + <i>B. phytofirmans</i> | 77.67 B | 12.67 AB | 21.67 B | 54.33 B |
| <i>S. ficaria</i> + <i>B. phytofirmans</i> | 75.00 B | 11.67 BC | 20.67 C | 49.00 D |
| <i>E. cloacae</i> + m <i>S. ficaria</i> × <i>B. phytofirmans</i> | 89.33 A | 14.00 A | 23.67 A | 59.33 A |

Different letters exhibit significant deference ($p \leq 0.05$) between treatments.

3.2 Yield Attributes

Multi-strain bacterization produced highly significant effects on 1000-grain weight of wheat, grain yield, straw yield and biological yield. Inoculation with *E. cloacae* + *S. ficaria* + *B. phytofirmans* remained significantly best among all the treatments regarding improvement in 1000-grains weight, grain, straw and biological yield. Inoculation with *E. cloacae* × *B. phytofirmans* remained significantly better over *E. cloacae* + *S. ficaria* for 1000-grains weight, grain, straw and biological yield (Tab. 3). In addition, no significant change was noted among effects of *E. cloacae* + *B. phytofirmans* and *S. ficaria* + *B. phytofirmans* regarding 1000-grain weight, grain, straw and biological yield. Single inoculation with *E. cloacae*, *S. ficaria* and *B. phytofirmans* also remained significantly better from control for 1000-grain, weight, grain, straw and biological yield. Maximum increases of 33, 71, 71 and 68% were noted in 1000-grains weight, grain, straw and biological yield respectively over control where *E. cloacae* + *S. ficaria* + *B. phytofirmans* was applied.

Table 3: Effect of single and multistrains inoculation of ACC deaminase producing PGPR on yield attributes of wheat

| Treatments | 1000 grains weight (g) | Grain Yield (Mg ha ⁻¹) | Straw Yield (Mg ha ⁻¹) | Biological Yield (Mg ha ⁻¹) |
|--|------------------------|------------------------------------|------------------------------------|---|
| Control | 45 F | 2.21 F | 6.62 F | 8.83 E |
| <i>E. cloacae</i> | 46 E | 2.94 CD | 8.81 CD | 11.7 C |
| <i>S. ficaria</i> | 51 D | 2.51 EF | 7.54 EF | 10.1 DE |
| <i>B. phytofirmans</i> | 51 D | 2.82 DE | 8.46 DE | 11.3 CD |
| <i>E. cloacae</i> + <i>S. ficaria</i> | 52 CD | 2.78 DE | 8.33 DE | 11.1 CD |
| <i>E. cloacae</i> + <i>B. phytofirmans</i> | 54 B | 3.35 B | 10.1 B | 13.4 AB |
| <i>S. ficaria</i> + <i>B. phytofirmans</i> | 53 BC | 3.32 BC | 9.97 BC | 13.3 B |
| <i>E. cloacae</i> + <i>S. ficaria</i> + <i>B. phytofirmans</i> | 60 A | 3.77 A | 11.3 A | 14.8 A |

Different letters exhibit significant deference ($p \leq 0.05$) between treatments.

3.3 Nutrients Concentration in Seeds and Shoot

Multi-strain PGPR inoculation significantly affected N, P and K concentration in shoot and seeds of wheat. The *E. cloacae* + *S. ficaria* + *B. phytofirmans* proved significantly better among all the treatments regarding N, P and K concentration in shoot and seeds. The *E. cloacae* + *B. phytofirmans* remained significantly better over *S. ficaria* + *B. phytofirmans* and exhibited significant increase in N, P and K concentration in shoot and seeds. The *S. ficaria* + *B. phytofirmans* remained better than *E. cloacae* + *S. ficaria* with respect to N concentration in seed and K in shoot. However, both the *S. ficaria* + *B. phytofirmans* and *E. cloacae* + *S. ficaria* remained statistically alike for N in shoot, K in seeds and P in seed and shoot (Tab. 4). Inoculation with *E. cloacae*, *S. ficaria* and *B. phytofirmans* in separate also proved significantly better from control for N, P and K concentration in shoot and seeds of wheat. Maximum increases of 37, 200, 57, 167, 28 and 38% were noted in N in seeds, N in shoot, P in seeds, P in shoot, K in seeds and K in shoot respectively over control where *E. cloacae* + *S. ficaria* + *B. phytofirmans* was applied.

Table 4: Effect of single and multistrains inoculation of ACC deaminase producing PGPR on nutrients concentration of wheat seed and shoot

| Treatments | N in seeds | N in shoot | P in seeds | P in shoot (%) | K in seeds | K in shoot |
|--|------------|------------|------------|----------------|------------|------------|
| Control | 1.35 G | 0.05 E | 0.30 G | 0.06 E | 1.80 F | 0.49 G |
| <i>E. cloacae</i> | 1.53 E | 0.07 DE | 0.37 E | 0.09 D | 2.20 D | 0.58 EF |
| <i>S. ficaria</i> | 1.44 F | 0.07 DE | 0.31 F | 0.10 CD | 2.10 E | 0.57 F |
| <i>B. phytofirmans</i> | 1.53 E | 0.08 D | 0.41 D | 0.11 C | 2.30 C | 0.59 DE |
| <i>E. cloacae</i> + <i>S. ficaria</i> | 1.64 D | 0.10 C | 0.43 C | 0.12 C | 2.30 C | 0.60 D |
| <i>E. cloacae</i> + <i>B. phytofirmans</i> | 1.72 B | 0.14 B | 0.45 B | 0.15 B | 2.40 B | 0.69 B |
| <i>S. ficaria</i> + <i>B. phytofirmans</i> | 1.65 C | 0.11 C | 0.43 C | 0.12 C | 2.30 C | 0.62 C |
| <i>E. cloacae</i> + <i>S. ficaria</i> + <i>B. phytofirmans</i> | 1.85 A | 0.15 A | 0.47 A | 0.16 A | 2.50 A | 0.79 A |

Different letters exhibit significant deference ($p \leq 0.05$) between treatments

4 Discussion

In the current study, the multi-strain PGPR inoculation significantly increased wheat plant height, spike length, number of spikelets spike⁻¹ and number of grains spike⁻¹ as compared to the control and even single-strain inoculation in most of the cases. The improvement in plant height might be due to reduction in stress ethylene level by sole consortium of ACC deaminase producing PGPR. Mayak et al. [31] stated that elevated 1-aminocyclopropane-1-carboxylic acid (ACC), especially under limited availability of water and nutrients in plant, increases ethylene concentration in root and shoot of plants. The plant roots and seeds exude the ACC into rhizosphere that is converted by ACC-deaminase into NH₃ and α -ketobutyrate thus decreasing ethylene level. Reduction in ethylene results in better roots elongation that might have facilitated plant to take in water and nutrients by exploring more soil volume [8]. In addition to ACC-deaminase activity, the improvement in wheat growth might also be favoured due to production of other growth-improving organic entities. Xie et al. [32] also noted IAA as an allied factor, playing a role in crops growth improvement. High IAA synthesis by PGPR increases surface area and length of adventitious and lateral roots in plants that help in nutrients uptake [33]. In addition, roots exudates (organic acids, phyto siderophores, sugars, vitamins, amino acids, nucleosides and mucilage) attract the PGPR for

colonization in roots with an improvement in the uptake of water and solubilization of immobilized (P and K) nutrients [34–35]. The multi-strains PGPR *E. cloacae* + *S. ficaria* + *B. phytofirmans* inoculation in specific, distinctly counteracted the adverse effects of drought in wheat in terms of improvement in 1000-grain weight, grain yield, straw yield and biological yield. This increase in yield of wheat might be due to an improvement in N, P and K uptake facilitated by the PGPR consortium. Plants growth is largely dependent on nutrients and H₂O holding potential of soil [10,37]. According to Hassan et al. [38], better uptake of N, P and K nutrients with PGPR application play a vital role in the development of shoot and root dry weight [36]. The probability of secretion of acid phosphatases enzyme may another important factor that would have helped in growth promotion through consortium [39].

5 Conclusion

Finally, the conclusion may be drawn out of the present study that multi-strain inoculation with the PGPR possessing an ACC-deaminase activity is more effective than single-strain inoculation to improve wheat growth and yield.

Conflicts of Interest: The authors declare no conflicts interests.

References

- Hodge, A. (2004). The plastic plant: root responses to heterogeneous supplies of nutrients. *New Phytologist*, 162(1), 9–24. DOI 10.1111/j.1469-8137.2004.01015.x.
- IFA (2015). International fertilizer industry association statistics. <http://www.fertilizer.org/ifadata>. accessed 18/3/2017.
- Goya, J. F., Frangi, J. L., Pérez, C., Dalla, T. F. (2008). Decomposition and nutrient release from leaf litter in Eucalyptus grandis plantations on three different soils in Entre Ríos, Argentina. *Bosque (Valdivia)*, 29(3), 217–226. DOI 10.4067/S0717-92002008000300005.
- Zhao, D., Reddy, K. R., Kakani, V., Read, J., Carter, G. (2003). Corn (*Zea mays* L.) growth, leaf pigment concentration, photosynthesis and leaf hyperspectral reflectance properties as affected by nitrogen supply. *Plant and Soil*, 257(1), 205–218. DOI 10.1023/A:1026233732507.
- Khan, M. I. R., Trivellini, A., Fatma, M., Masood, A., Francini, A. (2015). Role of ethylene in responses of plants to nitrogen availability. *Frontiers in Plant Science*, 6, 927.
- Hashmi, S., Younis, U., Danish, S., Munir, T. M. (2019). *Pongamia pinnata* L. leaves biochar increased growth and pigments syntheses in *Pisum sativum* L. exposed to nutritional stress. *Agriculture*, 9(7), 153. DOI 10.3390/agriculture9070153.
- Arshad, M., Frankenberger, W. T. Jr (2002). *Ethylene: agricultural sources and applications*. New York: Kluwer Academic Publishers.
- Glick, B. R. (1999). Enzymes that regulate ethylene levels-1-aminocyclopropane-1-carboxylic acid (ACC) deaminase, ACC synthase and ACC oxidase. *Indian Journal of Experimental Biology*, 35(1), 1–17.
- Yang, J., Kloepper, J. W., Ryu, C. M. (2008). Rhizosphere bacteria help plants tolerate abiotic stress. *Trends in Plant Science*, 14(1), 1–4. DOI 10.1016/j.tplants.2008.10.004.
- Glick, B. R. (1995). The enhancement of plant-growth by free-living bacteria. *Canadian Journal of Microbiology*, 41(2), 109–117. DOI 10.1139/m95-015.
- Glick, B. R. (2005). Modulation of plant ethylene levels by the bacterial enzyme ACC deaminase. *FEMS Microbiology Letters*, 251(1), 1–7. DOI 10.1016/j.femsle.2005.07.030.
- Pinton, R., Varanini, Z., Naniperi, P. (2001). The rhizosphere as a site of biochemical interactions among soil components, plants and microorganisms. In: *The Rhizosphere*. CRC Press, Boca Raton, 17–34.
- Kloepper, J. W., Lifshitz, R., Zablotowicz, R. M. (1989). Free-living bacterial inocula for enhancing crop productivity. *Trends in Biotechnology*, 7(2), 39–44. DOI 10.1016/0167-7799(89)90057-7.

14. Ngumbi, E., Klopper, J. (2016). Bacterial-mediated drought tolerance: current and future prospects. *Applied Soil Ecology*, 105, 109–125. DOI 10.1016/j.apsoil.2016.04.009.
15. Shakir, M. A., Bano, A., Arshad, M. (2012). Rhizosphere bacteria containing ACC-deaminase conferred drought tolerance in wheat grown under semi-arid climate. *Soil & Environment*, 31, 108–112.
16. Cherif, H., Marasco, R., Rolli, E., Ferjani, R., Fusi, M. et al. (2015). Oasis desert farming selects environment-specific date palm root endophytic communities and cultivable bacteria that promote resistance to drought. *Environmental Microbiology Reports*, 7(4), 668–678. DOI 10.1111/1758-2229.12304.
17. Karthikeyan, S., Zhou, Q., Zhao, Z., Kao, C., Tao, Z. et al. (2004). Structural analysis of Pseudomonas 1-aminocyclopropane-1-carboxylate deaminase complexes insight into the mechanism of a unique pyridoxal-5-phosphat dependent cyclopropane ring opening reaction. *Biochemistry*, 43(42), 13328–13339. DOI 10.1021/bi048878g.
18. Danish, S., Zafar-ul-Hye, M., Hussain, S., Riaz, M., Qayyum, M. F. (2019). Mitigation of drought stress in maize through inoculation with drought tolerant ACC deaminase containing PGPR under axenic conditions. *Pakistan Journal of Botany*, 52(1). DOI 10.30848/PJB2020-1(7)
19. Danish, S., Zafar-ul-Hye, M. (2019). Co-application of ACC-deaminase producing PGPR and timber-waste biochar improves pigments formation, growth and yield of wheat under drought stress. *Scientific Reports*, 9(1), 5999. DOI 10.1038/s41598-019-42374-9.
20. Henderson, K. N., Tye-Din, J. A., Reid, H. H. (2007). A structural and immunological basis for the role of human leukocyte antigen DQ8 in celiac disease. *Immunity*, 27(1), 23–34. DOI 10.1016/j.immuni.2007.05.015.
21. Pakistan Economic Survey (2019). *Agriculture. Chapter 2*. Ministry of Finance, 1–33.
22. Moodie, C. D., Smith, H. W., McCreery, R. A. (1959). *Laboratory manual for soil fertility*. Washington: Department of Agronomy, State College of Washington Pullman, 31–39.
23. Olsen, S. R., Cole, C. V., Watanabe, F. S., Dean, L. A. (1954). Estimation of available phosphorus in soils by extraction with sodium bicarbonate. US Department of Agriculture Circular No. 939. Banderis, A. D., Barter, D. H., Anderson, K. Agricultural and Advisor.
24. Nadeem, S. M., Zahir, Z. A., Naveed, M., Nawaz, S. (2013). Mitigation of salinity-induced negative impact on the growth and yield of wheat by plant growth-promoting rhizobacteria in naturally saline conditions. *Annals of Microbiology*, 63(1), 225–232. DOI 10.1007/s13213-012-0465-0.
25. Walkley, A. (1935). An examination of methods for determining organic carbon and nitrogen in soils. *Journal of Agricultural Science*, 25(4), 598–609. DOI 10.1017/S0021859600019687.
26. Dworkin, M., Foster, JW. (1957). Studies on Pseudomonas methanica (Söhngen) nov. comb. *Journal of Bacteriology*, 72(5), 646–659. DOI 10.1128/JB.72.5.646-659.1956.
27. Wolf, B. (1982). The comprehensive system of leaf analysis and its use for diagnosing crop nutrient status. *Communications in Soil Science and Plant Analysis*, 3(12), 1035–1059. DOI 10.1080/00103628209367332.
28. Jones, J. B. Jr., Wolf, B., Mills, H. A. (1991). Athens: Micro-Macro Publishing Inc., Athens, Georgia, USA.
29. Jackson, M. L. (1962). *Soil chemical analysis*. Englewood Cliffs: Prentice Hall.
30. Steel, R. G. D., Torrie, J. H. (1997). *Principles and procedures of statistics-a biometric approach*. Third Edition. Singapore: McGraw Hill Book International Co., 204–227.
31. Mayak, S., Tirosh, T., Glick, B. R. (2004). Plant growth-promoting bacteria confer resistance in tomato plants to salt stress. *Plant Physiology and Biochemistry*, 42(6), 565–572. DOI 10.1016/j.plaphy.2004.05.009.
32. Xie, H., Pasternak, J. J., Glick, B. R. (1996). Isolation and characterization of mutants of the plant growth-promoting rhizobacterium *Pseudomonas putida* gr12-2 that overproduce indoleacetic acid. *Current Microbiology*, 32(2), 67–71. DOI 10.1007/s002849900012.
33. Mohite, B. (2013). Isolation and characterization of indole acetic acid (IAA) producing bacteria from rhizospheric soil and its effect on plant growth. *Journal of Soil Science and Plant Nutrition*, 13, 638–649.
34. Basak, B. B., Biswas, D. R. (2010). Co-inoculation of potassium solubilizing and nitrogen fixing bacteria on solubilization of waste mica and their effect on growth promotion and nutrient acquisition by a forage crop. *Biology and Fertility of Soils*, 46(6), 641–648. DOI 10.1007/s00374-010-0456-x.

35. Drogue, B., Combes-Meynet, E., Moëgne-Loccoz, Y., Wisniewski-Dyé, F., Prigent-Combaret, C. (2013). Control of the cooperation between plant growth-promoting rhizobacteria and crops by rhizosphere signals. In: de Bruijn, F. J., ed. *Molecular Microbial Ecology of the Rhizosphere*. NJ, USA: John Wiley & Sons, Inc., 281–294.
36. Hassan, W., Bashir, S., Ali, F., Ijaz, M., Hussain, M. et al. (2016). Role of ACC-deaminase and/or nitrogen fixing rhizobacteria in growth promotion of wheat (*Triticum aestivum* L.) under cadmium pollution. *Environmental Earth Sciences*, 75(3), 267. DOI 10.1007/s12665-015-4902-9.
37. Kloepper, J. W., Schroth, M. N. (1978). Plant growth promoting rhizobacteria on radishes. *Proceedings of the 4th International Conference on Plant Pathogenic Bacteria*, Angers, France, pp. 879–882.
38. Hassan, W., Hussain, M., Bashir, S., Shah, A. N., Bano, R. et al. (2015). ACC-deaminase and/or nitrogen fixing rhizobacteria and growth of wheat (*Triticum aestivum* L.). *Journal of Soil Science and Plant Nutrition*, 15, 232–248.
39. Dodor, D. E., Tabatabai, M. A. (2003). Effect of cropping systems on phosphatases in soils. *Journal of Plant Nutrition and Soil Science*, 166(1), 7–13. DOI 10.1002/jpln.200390016.
40. Zafar-ul-Hye, M., Danish, S., Abbas, M., Ahmad, M., Munir, T. (2019). ACC deaminase producing PGPR *Bacillus amyloliquefaciens* and *Agrobacterium fabrum* along with biochar improve wheat productivity under drought stress. *Agronomy*, 9(7), 343. DOI 10.3390/agronomy9070343.