

# Growth of Non-enzymatic Cholesterol Biosensor using TiO<sub>2</sub> Decorated Graphene Oxide with Bare GCE and PPy-GCE

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## ABSTRACT

*The cholesterol level determination is a significant clinical diagnostic solution for heart and thrombosis problems. In this work, we examined a novel non-enzymatic cholesterol biosensor using cholesterol oxidase (ChOx) enzyme immobilized on TiO<sub>2</sub> nanoparticles influenced by reduced graphene oxide (rGO) - polypyrrole (PPy) (rGO-TiO<sub>2</sub>/PPy-GCE) nanocomposite was developed on a glassy carbon electrode (GCE) and the higher sensing response with lower detection limits were observed. The electrochemical properties of GCE modified PPy (PPy-GCE) were studied using CV (Cyclic Voltammetry) and DPV (Differential Pulse Voltammetry). The reported sensor exhibited piecewise linearity in the range of 0.1 μM to 1 μM and 1 μM to 600 μM with the sensing response of 61.665 and 0.1466 (2μA μM×cm) respectively. The detection limits of the sensor were found to be 32 nm. The results were repeatable and reproducible and this sensor can be applied to determine the cholesterol at real sample with satisfactory results.*

**KEYWORDS:** *Clinical analysis, Non-enzymatic, Nanomaterials, Chemical oxidative polymerisation*

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## 1. INTRODUCTION

Cholesterol and other fatty acids are important components of the human body. Cholesterol helps the body make cell membranes, many hormones, and vitamin D. Cholesterol comes from food and the liver. Cholesterol and other fats are carried in the bloodstream as spherical particles (ie., lipoproteins)<sup>[1]</sup>. The normal total cholesterol level in healthy human serum is approximately 200 mg dl (or 5.17 mM)<sup>[2, 3]</sup>. Estimation of cholesterol levels is extremely important for clinical diagnosis. Different approaches, including high performance liquid chromatography<sup>[4-7]</sup>, near infrared (IR) spectra<sup>[8-10]</sup> and colorimetric assay<sup>[11-13]</sup>, have been used to detect cholesterol levels. However, the above approaches are expensive and time consuming and require large amounts of serum samples<sup>[14]</sup>. Therefore, a rapid, robust and sensitive cholesterol detection method must be developed.

Among these methods, electrochemical detection has attracted and considerable attention because of its speed, simplicity, and low cost. Most of the literature on electrochemical based cholesterol sensors has focused on determination based on the activity of the enzyme "cholesterol oxidase"<sup>[15]</sup>. Alagappa et al.,<sup>[16]</sup> reported an electrochemical cholesterol biosensor based on cholesterol oxidase (ChOx) enzyme immobilized on gold nanoparticles – functionalized – multiwalled carbon nanotube (MWCNT) – polypyrrole (PPy) nanocomposite modified electrode. The sensor was fabricated by a two-step approach wherein the Au NPs-f-MWCNT was prepared by wet chemical method followed by electro polymerization of pyrrole. The AuNPs-f-MWCNT-PPy-ChOx/GCE showed a linear

response from  $2 \times 10^{-3}$  to  $8 \times 10^{-3}$ /M in amperometry with a sensitivity and detection limit of  $10.12/\mu\text{A}/\text{mM}^{-1} \text{cm}^{-3}$  and  $0.1 \times 10^{-3}$ /M respectively. Ansari et al.<sup>[17]</sup> studied the CHIT–tin oxide ( $\text{SnO}_2$ ) nano-biocomposite film for cholesterol biosensor development. They showed that ChOx/CHIT- $\text{SnO}_2$ /ITO (Indium–Tin–Oxide) was more stable than the ChOx/CHIT/ITO bioelectrode, and it presented a high sensitivity of  $34.7\text{mA}/\text{mgdL}^{-1}\text{cm}^2$ , a linear response in the range of 10–400mg/dL, and a low detection limit of 5mg/dL. Nguyet et al.<sup>[18]</sup> reported a DNA sensor based on the  $\text{CeO}_2$ /PPy nanocomposite for Salmonella detection. In situ chemical oxidative polymerization was used to prepare the core–shell  $\text{CeO}_2$ -NR@PPy nanocomposite, which provided a suitable platform for electrochemical DNA biosensor fabrication. The sDNA/ $\text{CeO}_2$ NRs@PPy/electrode response under optimal conditions presented a linearity between 0.01 and 0.4nM with a sensitivity of  $593.7\Omega \cdot \text{nM}^{-1} \cdot \text{cm}^{-2}$ . Tiwari and Gong<sup>[19]</sup> reported a novel chitosan (CHIT)/ $\text{SiO}_2$ /multiwalled carbon nanotube (MWCNT)/electrode-based cholesterol biosensor. They established the linear relationship between oxidation current response and cholesterol concentrations to be in the range of 5.0–5000  $\mu\text{g}/\text{mL}$ , a response time of 5s, and a sensitivity of  $3.4\text{nA}/\text{mg dL}^{-1}$ . Wisitsoraat et al.,<sup>[20]</sup> developed a new cholesterol biosensor by using carbon nanotubes (CNT) directly growth on a glass-based chip via the low-temperature chemical vapor deposition process. They determined the linear detection range to be between 50 and 400mg/dL with a sensitivity of  $0.0512\text{nA}/\text{mg} \cdot \text{dL}^{-1}$ .

Among the metal oxide NPs based biosensor, the interest for titanium dioxide ( $\text{TiO}_2$ ) NPs is

being attention due to its unique properties such as high surface area and high catalytic efficiency, which can improve the interaction between biomolecules and electrode surfaces [21–25]. Recently, cholesterol bio-sensor based on TiO<sub>2</sub> nanotubes decorated with Cu<sub>2</sub>O nanoparticles has been reported [26]. Particularly, reduced graphene oxide performed cholesterol oxidase (ChOx) exhibits fast response and repeatable cyclic process in sensor [27] and also the polypyrrole (PPy) influenced electrochemical sensor shows flexibility, better electrical conductivity and surface characteristics [28]. Meanwhile, electrode influences is the important parameter in sensor, a lot of work were mentioned [16–20] with different electrode but the higher efficiency with fast responded electrode of glassy carbon electrode was not preferred. The merits of the biosensor using TiO<sub>2</sub> nanoparticles, reduced graphene oxide (rGO), polypyrrole (PPy) developed on glassy carbon electrode (GCE) was reported as many but the compositional work using glassy carbon electrode (GCE) and their sensing response was not reported on literature survey.

In the present work, a novel biosensor based on the cholesterol oxidase (ChOx) enzyme immobilized on TiO<sub>2</sub> nanoparticles influenced by reduced graphene oxide (rGO)/polypyrrole (PPy) (rGO-TiO<sub>2</sub>/PPy-GCE) nanocomposite was developed on a glassy carbon electrode (GCE).

## **2. EXPERIMENTAL AND MATERIAL CHARACTERIZATION**

### **2.1 Reagents and Sample Preparation**

All the chemicals were used analytical grade.

Cholesterol, Triton X-100 & IPA, diethyl ether, petroleum, hydrogen peroxide, potassium permanganate, sulphuric acid, hydrochloric acid, potassium ferricyanide (K<sub>3</sub>[Fe(CN)<sub>6</sub>]), sodium hydroxide pellets, potassium dihydrogen orthophosphate (KH<sub>2</sub>PO<sub>4</sub>) and acetone were bought from Loba chemie. Ethanol was brought from Changshu Yanguan Chemical Ltd., Ultrapure water (MilliQ, Millipore water) was used for all the experiments. Since cholesterol is not soluble in aqueous media, stock solutions were prepared by dissolving cholesterol initially in Triton X-100 and then suitable solvents (0.1 M NaOH or PBS) were added. The composition of the solutions used will be explained in the respective sections.

Graphene oxide (GO) was synthesized directly from graphite by the Hummers method. The rGO and rGO-TiO<sub>2</sub> nanocomposites synthesis was assisted by hydrothermal process. Nanocomposites rGO-TiO<sub>2</sub>/PPy was synthesized by in-situ chemical oxidative polymerization technique. A bare glassy carbon electrode (GCE) or a modified GCE was applied as the working electrode, meanwhile, a saturated calomel electrode (SCE) and a platinum wire were used as the reference and counter electrodes, respectively.

The aliquot of cholesterol was extracted from egg yolk for the real sample analysis to evaluate the sensor performance. The yellow yolk portion of egg is removed separately, washed and filtered to remove other contents. A small puncture was made in the slaggy yolk and pure yolk was taken for the further electrochemical analysis. 10 g of egg yolk was weighed and then diluted to 50 mL using distilled water. 5 mL of aliquot was mixed thoroughly with 5 mL of IPA, 12.5 mL diethyl ether and 12.5 mL of petroleum ether in a separating funnel. The mixture was shaken well and the inorganic layer was separated from the organic layer. The organic layer was separated and the organic solvents were allowed to evaporate.

### **2.2 Material Characterization**

The prepared samples were characterized by Scanning electron microscopy (SEM, Hitachi S-4800), Fourier transform infrared spectroscopy (FT-IR-8700 spectrometer), UV-Visible spectroscopy (JASCO V-530) and Thermal gravimetric analysis (NETZSCH STA 2500).

All electrochemical experiments were performed on a CHI 920C electrochemical workstation (SREC, Coimbatore) with a conventional three-electrode system. The electrode was standardized by performing Cyclic voltammetry (CV) in 1 mM of potassium ferricyanide ( $K_3[Fe(CN)_6]$ ) and 100 mM of potassium chloride (KCl) as supporting electrolyte with initial and final potential of -0.25 V to +0.5 V respectively and the scan rate of 50 mV/s. Ferulic acid was sensed using CV and differential pulse voltammetry (DPV) with the presence of some concentration of ferulic acid in phosphate buffer. The CV parameters for detection of ferulic acid were -0.3 V to 0.9 V with the scan rate of 50 mV/s, the DPV parameters were 0 to 1.5 V as initial and final potential, 0.05 V as pulse increment. The electrode was modified and standardized after each electrochemical measurement.

### 3. RESULTS AND DISCUSSIONS

#### 3.1 Functional Group Analysis of GO, rGO and rGO-TiO<sub>2</sub>

Graphene oxide (GO), reduced graphene oxide (rGO) and TiO<sub>2</sub> decorated graphene oxide (rGO-TiO<sub>2</sub>) were characterized for their presence of functional groups using FT-IR spectroscopy. Fig. 1 shows the obtained FT-IR spectra for GO, rGO and rGO-TiO<sub>2</sub>. The absorption band near 522 cm<sup>-1</sup> corresponded to the Ti-O vibration<sup>[29]</sup>. The peak strength of rGO-TiO<sub>2</sub> nanocomposites is lower than GO and some of the peaks even disappear. From the FT-IR spectrum of rGO-TiO<sub>2</sub> nanocomposites, few peaks indicating the

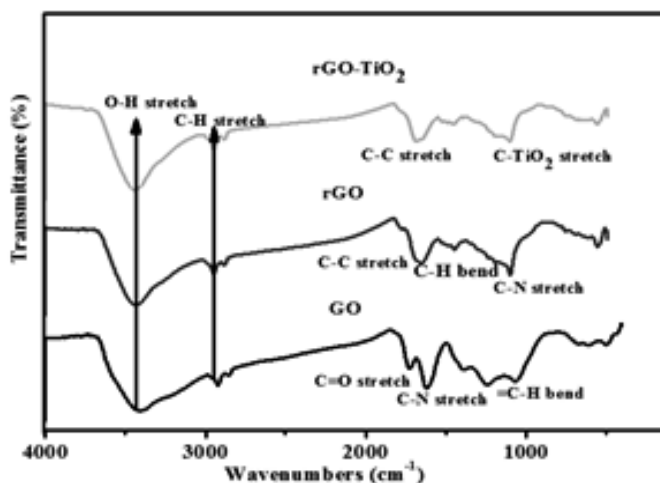


Figure 1. FT-IR spectra of GO, rGO and rGO-TiO<sub>2</sub>

oxygen containing functional groups can be seen after reduction. These oxygen-containing functional groups demonstrate the successful preparation of GO and provide anchoring sites for the adsorption of heavy metal ions on the TiO<sub>2</sub>/GO-8 nanocomposites<sup>[30]</sup>. The FT-IR spectra of raw and coated fabrics displayed a

broad absorption band of around 3285cm<sup>-1</sup>, which is attributed to the O-H stretching vibration of C-OH groups<sup>[31]</sup>. GO exhibited functional groups like -OH, -COOH, C=O whereas rGO exhibited reduced levels of -OH and negligible amount of C=O stretch which results in enhanced conductivity. The spectra

of  $\text{rGO-TiO}_2$  exhibit the presence of  $\text{TiO}_2$  in the form of  $\text{C-TiO}_2$  functional group which confirmed the formation of  $\text{TiO}_2$  nanoparticles.

### 3.2 Absorption Spectra of GO, rGO and $\text{rGO-TiO}_2$

UV-Vis spectra of GO, rGO and  $\text{rGO-TiO}_2$  is presented in Fig. 2. UV-Vis spectrum of GO exhibits the absorption at 230 nm which is due to  $\text{p-p}^*$  transition<sup>[32]</sup>. The spectrum for rGO

exhibits the absorption at the shifted wavelength of 260 nm compared to GO. Fig shows  $\text{rGO-TiO}_2$  exhibits the absorption for rGO at 260 nm and an another absorption peak at 350 nm which corresponds to the presence of  $\text{TiO}_2$ <sup>[33]</sup>. The absorption of reduced graphene shifts from 226 to 256 nm, suggesting that the electronic conjugation within graphene sheets is restored after the reduction<sup>[34]</sup>.

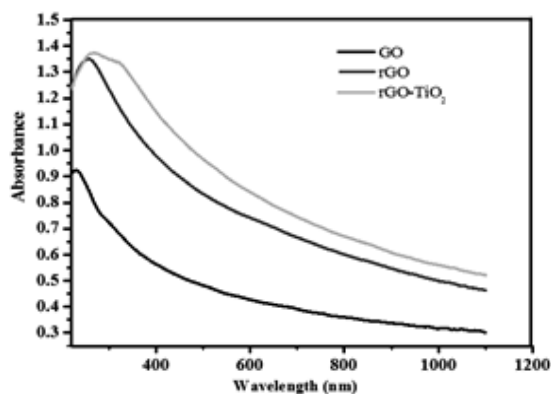


Figure 2. UV-visible spectrum for GO, rGO,  $\text{rGO-TiO}_2$

### 3.3 Thermo Gravimetric Analysis of GO, rGO and $\text{rGO-TiO}_2$

Thermo gravimetric analysis (TGA) helps to know the weight decomposition of the sample with

respect to the increase in temperature. TGA plot (Fig. 3) gives the information about the amount of the material loss with respect to the temperature.

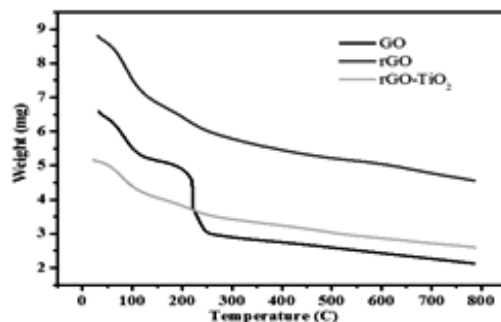


Figure 3. TGA analysis of GO, rGO and  $\text{rGO-TiO}_2$

GO sample taken for TGA was of weight 6.5870 mg where with increase in temperature, at temperature 210°C there is decomposition occurring amounting to 54.90 % of loss which is equivalent to 3.616 mg and from temperatures 260°C to 800°C the decomposition amounts to 12.80% which is 0.8429 mg of weight. TGA plot of rGO exhibits the decomposition of 48.27% with the weight

loss of 4.29 mg from the temperature 50°C to 800°C. TGA plot of rGO-TiO<sub>2</sub> exhibits the decomposition of 49.64% with the weight loss of 2.567 mg from 50°C to 800°C.

### 3.4 Surface Analysis of rGO-TiO<sub>2</sub>

The surface morphology and topology of rGO-TiO<sub>2</sub> nanocomposite was characterized using SEM. Fig. 4 shows the clusters of TiO<sub>2</sub>

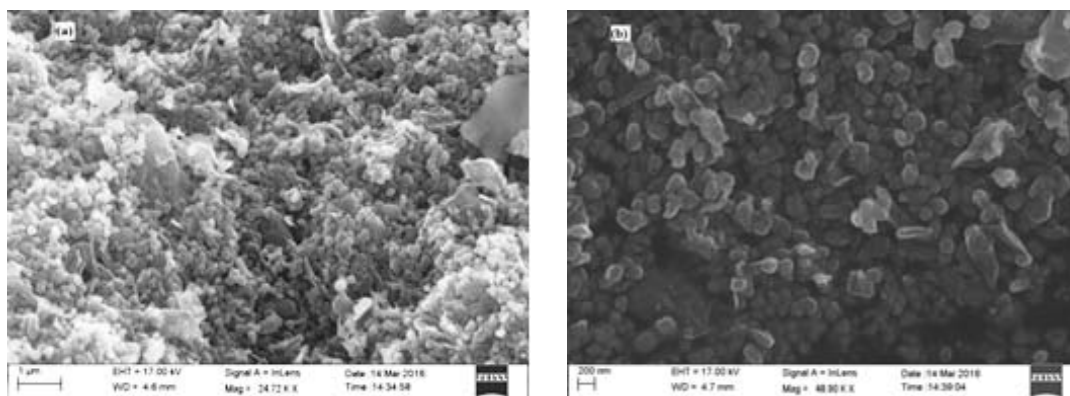


Figure 4. SEM image of (a) 1 μm and (b) 200 nm rGO-TiO<sub>2</sub>

nanoparticles decorated on the reduced graphene sheets. The formation of the nanoparticles is merely spherical in shape with the size of approximately 135 nm. Thus it is confirmed that TiO<sub>2</sub> decorated rGO has enhanced surface area and enhanced conductivity.

### 3.5 Calculation of Effective Surface Area

Electrochemically active area of modified electrode was calculated by performing chronocoulometry for K<sub>3</sub>[Fe(CN)<sub>6</sub>]. The Cottrell charge plot for bare electrode and modified electrode is shown in Fig. 5 which confirms the increase in electrochemically active surface area. The surface area is calculated using Cottrell equation.

$$\text{Slope} = \frac{2nFAC\sqrt{D}}{\sqrt{\pi}}$$

By substituting the known values, the surface area of the PPy modified GCE was 0.262\*10<sup>-4</sup> m<sup>2</sup> which is enhanced compared to the surface area of bare GCE which was of 0.196 \*10<sup>-4</sup>m<sup>2</sup>. The diffusion co-efficient for bare GCE was 0.430\*10<sup>-2</sup> m<sup>2</sup>/s.

### 3.6 Catalytic Activity of Modified Electrode

The catalytic activity of modified electrode was exhibited by performing Cyclic voltammetry with 1mM of K<sub>3</sub>[Fe(CN)<sub>6</sub>] as redox species and 100 mM of KCl as the supporting electrolyte.

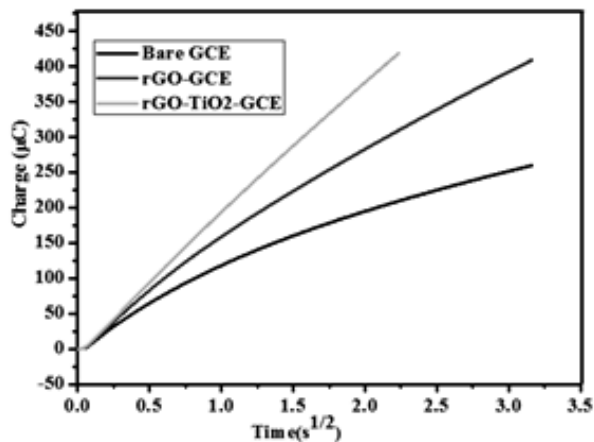


Figure 5. Chronoamperometry for bare GCE, rGO/GCE and rGO-TiO<sub>2</sub>/GCE

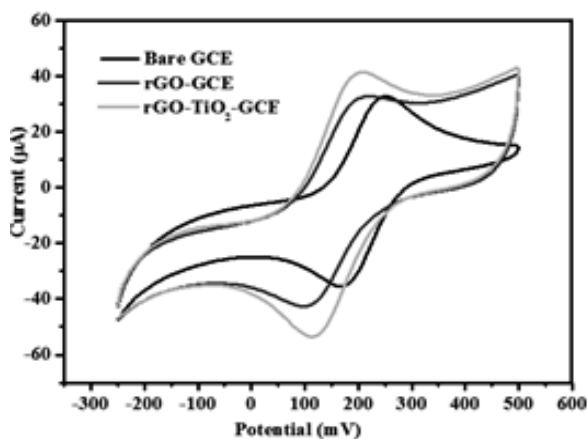


Figure 6. Cyclic Voltammogram for bare GCE, rGO/GCE, rGO-TiO<sub>2</sub>/GCE

On performing CV for potassium ferricyanide, it results in one electron redox transfer Fig. 6 exhibits redox curve for potassium ferricyanide, the oxidation occurred at +259 mV and reduction occurred at +168 mV. In order to get better conductivity, the catalytic activity of the PPy-GCE is shown in Fig. 6 where there is increase in current and also left shift in reduction and oxidation potential compared to bare GCE. This confirms the

enhanced surface area for modified electrode which helps in the increase of conductivity and sensitivity of the electrode. The peak separation potential for redox species on bare GCE was 88 mV whereas for rGO/PPy-GCE it was 95 mV and for rGO-TiO<sub>2</sub>/PPy-GCE it was 86 mV. Catalytic activity of bare GCE, rGO/PPy-GCE and rGO-TiO<sub>2</sub>/PPy-GCE in terms of anodic potential, cathodic potential and area under the curve are shown in the table.1

TABLE 1. Catalytic activity of bare GCE, rGO/PPy-GCE and rGO-TiO<sub>2</sub>/PPy-GCE in terms of anodic and cathodic potential and area under the curve

Electrode	Area under the curve		Peak potential	
	Cathodic	Anodic	Anodic	Cathodic
Bare GCE	5.901 $\mu$ W	15.31 $\mu$ W	165 mV	253 mV
rGO/PPy-GCE	10.912 $\mu$ W	11.620 $\mu$ W	114 mV	209 mV
rGO-TiO <sub>2</sub> /PPy-GCE	12.494 $\mu$ W	14.057 $\mu$ W	116 mV	202 mV

### 3.7 Optimization

Buffer used for the detection and quantification of cholesterol is phosphate buffer. The effect of pH in the buffer is studied by varying pH from 4.5 to 11.5.

Fig. 7 gives the effect of pH in the detection of cholesterol. It is evident that the pH of 7.45 is more suitable for the detection of cholesterol with maximum current response compared to other pH values.

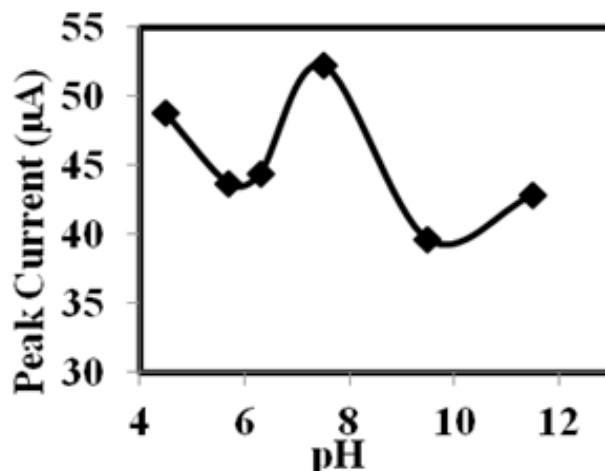


Figure 7. Calibration plot for the detection of cholesterol with various phosphate buffer

### 3.8 Scan Rate Variation and Detection of Cholesterol

CV was performed by varying the scan rates for the detection of cholesterol from 1 mV/s to 500 mV/s. From Fig. 8 it is observed that with increase in scan rate, there is linear increase in current response. The detection of cholesterol

on bare GCE and rGO-TiO<sub>2</sub>/PPy-GCE was done using cyclic voltammetry. Fig. 9 exhibits the cyclic voltammogram of 100  $\mu$ M cholesterol and the oxidation peak was observed at 0.685 V for bare GCE and 0.619 V for rGO-TiO<sub>2</sub>/PPy-GCE. This shows that the modified electrode shows better sensitivity and conductivity for the detection of cholesterol.



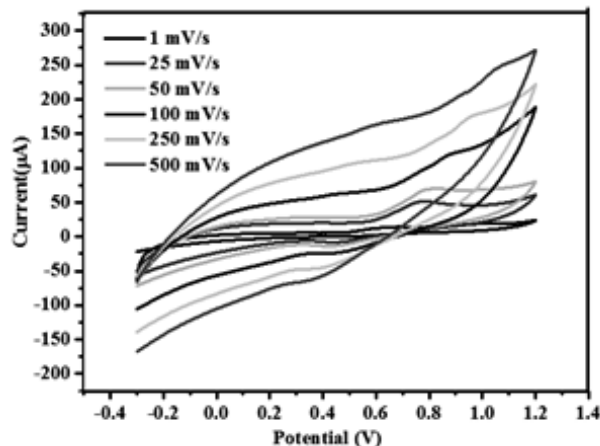


Figure 8. Cyclic voltammograms for detection of 100  $\mu\text{M}$  cholesterol in optimized phosphate buffer

### 3.9 Linear Range and Limit of Detection

In order to detect the cholesterol of lower concentrations with high sensitivity and differential pulse voltammetry (DPV) was used as a technique for detection of cholesterol. The DPV parameters were optimized such that sharp oxidation peak was obtained with the presence of ferulic acid.

Figure 10 shows the differential pulse voltammograms for various concentrations from 0.1  $\mu\text{M}$  to 600  $\mu\text{M}$ . The PPy modified GCE exhibited piecewise linear range for the cholesterol detection from 0.1  $\mu\text{M}$  to 1  $\mu\text{M}$  ( $y = 12.108x + 25.71$ ,  $R^2 = 0.9908$ ) and 1  $\mu\text{M}$  to 600  $\mu\text{M}$  ( $y = 0.0288x + 39.049$ ,  $R^2 = 0.9798$ ) (Fig. 11). The sensitivity was calculated for 0.1  $\mu\text{M}$  to 1  $\mu\text{M}$  and 1  $\mu\text{M}$  to 600  $\mu\text{M}$  was 61.665 and 0.1466 respectively. The limit of quantification (LoQ) of Cholesterol for the rGO- $\text{TiO}_2$ /PPy-GCE was found to be 0.1  $\mu\text{M}$ . The limit of detection (LoD) of Cholesterol for rGO- $\text{TiO}_2$ /PPy-GCE was found to be 0.05  $\mu\text{M}$ . From the literature of electrochemical detection of

cholesterol, various materials were applied for the detection. As far as our knowledge, metal oxide decorated reduced graphene oxide was not employed in the detection of Cholesterol. By comparing the common sensor characteristics like linear range and limit of detection, the present method gives better response which is listed in Table 2.

### 4.0 REPEATABILITY, REPRODUCIBILITY AND SELECTIVITY

The developed sensor was checked for the repeatability, reproducibility and the results were repeatable and reproducible with the deviation of  $\pm 2.78\%$  and  $\pm 2.21\%$  respectively. The sensor was tested for the selectivity with the presence of interferences like Uric acid, dopamine and ascorbic acid, the developed sensor is observed to be selective for cholesterol. The developed sensor was employed in detecting cholesterol in food samples like commercially available vanilla essence, vanilla flavored biscuit, vanilla flavored ice-cream and vanilla flavored cake mix. The detection was

TABLE 2. Comparison of different electrochemical methods for detection of Cholesterol

Reference	Electrode type	Limit of detection ( $\mu\text{M}$ )	Linear range ( $\mu\text{M}$ )
[35]	G/PVP/PANI nanocomposites	1	0.05-10
[36]	Graphene/GCE	0.056	0.6-48
[37]	MWCNT-PPy	1	0.4-6.5
[38]	Screen printed electrode	0.4	5-400
[39]	AuPd NP/Graphene/GCE	0.02	0.1-7 and 10-40
[40]	Au-f-MWCNT-PPy	2.88	2-8
[41]	AgNPs/Graphene nanosheets/GCE	0.332	2-100
[42]	Anode pre-treated BDDE	0.16	3.3-98
Present Work	rGO-TiO <sub>2</sub> /PPy-GCE	0.05	0.1-1 and 1-600

GCE-Glassy carbon electrode, AuPd NP- Gold Palladium Nanoparticles, CPE-Carbon paste electrode, Ag NPs- Silver Nanoparticles, poly (vinyl pyrrolidone) (PVP), polyaniline (PANI) multiwalled carbon nanotubes (MWCNT) and Polypyrrole (PPy).

successfully done and the results were validated using UV-Vis spectroscopy. The results are listed in Table 3.

## 5. CONCLUSION

A PPy modified Glassy carbon electrode was employed in detection of cholesterol. The

TABLE 3. Real sample results with validation results using UV-Vis spectroscopy

Sample	Electrochemical		UV	Recovery (%)		
	$\mu\text{g/mL}$	RSD (%)	$\mu\text{g/mL}$	RSD(%)		
Biscuit	9.419	1.95	8.852	1.09	93.98	106.40
Pudding powder	35.511	1.71	29.95	0.606	84.34	118.56
Vanilla essence	39.703	1.84	36.271	2.42	91.355	108.12
Ice cream	10.122	3.75	12.13	1.37	119	83.36

modified electrode exhibited an excellent catalytic activity because of the presence of metal oxide. Selective and sensitive detection of cholesterol was obtained with wide linear range, low detective limit. The developed sensor was successfully applied in detection of cholesterol in food samples like vanilla essence, vanilla flavored biscuits, ice cream and cake

mix. The electrochemical results were validated using UV-visible spectrometry. The sensitivity was calculated for 0.1  $\mu\text{M}$  to 1  $\mu\text{M}$  and 1  $\mu\text{M}$  to 600  $\mu\text{M}$  was 61.665 and 0.1466 respectively.

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