

MicroRNA-520b Suppresses Proliferation, Migration, and Invasion of Spinal Osteosarcoma Cells via Downregulation of Frizzled-8

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Spinal osteosarcoma (OS) is a malignant tumor that has a poor outcome. MicroRNA-520b (miR-520b) acts as a cancer suppressor in various types of cancer. Because of the limited amount of literature on OS, we aimed to identify the role of miR-520b in OS. The miR-520b level in clinical spinal OS tissues and adjacent nontumor tissues as well as in cell lines was assessed. The effect of miR-520b on cell proliferation, migration, invasion, and frizzled-8 (FZD8) degradation were all evaluated. Alterations of key proteins involved in the Wnt/ β -catenin pathway were assessed by Western blot analysis. In the present study, miR-520b was downregulated in human spinal OS tissues and OS cell lines ($p < 0.01$ or $p < 0.001$). Overexpression of miR-520b inhibited cell proliferation ($p < 0.01$ or $p < 0.001$), migration ($p < 0.01$), and invasion ($p < 0.01$). FZD8 expression was negatively regulated by infection with a lentivirus vector carrying an miR-520b precursor in dose- and time-dependent manners. In OS tissues, miR-520b was inversely correlated with FZD8 expression. FZD8 was upregulated in human spinal OS tissues and cell lines. Finally, miR-520b inactivated the Wnt/ β -catenin pathway through downregulation of FZD8. miR-520b inhibited cell proliferation, migration, and invasion through inactivating the Wnt/ β -catenin pathway by downregulation of FZD8, providing a novel therapeutic target for spinal OS.

Key words: miR-520b; Spinal osteosarcoma (OS); Frizzled-8; Wnt/ β -catenin pathway

INTRODUCTION

Osteosarcoma (OS) is the most common primary bone cancer and the third most common malignant tumor in children and adolescents^{1,2}. In this age group, OS accounts for 2%–3% of all malignancies, resulting in an average incidence of 7.7 per million worldwide^{3,4}. OS is reported to originate from mesenchymal stem cells and presents as a production of an osteoid matrix by the neoplastic cells⁵. Commonly, OS occurs in the metaphysis of the long bone while rarely occurring in the spine, which only accounts for 3%–5% of all spinal malignancies^{6,7}. Clinical study shows that OS troubles patients with pain and swelling in the affected bone, as well as creating a neurologic deficit in approximately 70% of all OS patients^{8,9}. At the molecular level, genetic disorders are greatly associated with OS⁶.

MicroRNAs (miRs) are a family of small noncoding RNAs that transcriptionally or posttranscriptionally modulate gene expression by suppressing the transcription or degrading the mRNA of target genes¹⁰. Accumulating evidence has proven that miRs participate in the regulation of OS progression. Among those miRs, some act as potential tumor suppressors in OS, such as miR-454¹¹ and

miR-217¹², whereas some act as oncogenes in OS, such as miR-374a¹³ and miR-664¹⁴. miR-520b is a recently recognized tumor suppressor that has been verified in an array of cancer types. A previous study suggested that miR-520b could suppress cell proliferation of hepatoma cells¹⁵. Another study also reported that miR-520b inhibited cell proliferation and migration in gastric cancer¹⁶. Furthermore, miR-520b enhances the sensibility of breast cancer cells to complement attack¹⁷. However, few investigations identified the specific role of miR-520b in OS progression. Whether miR-520 acts as a cancer suppressor in OS remains unclear.

Currently, the strategy for OS is a combination of neoadjuvant chemotherapy and surgical resection¹⁸. Although modern medicine for OS significantly improves the 5-year survival rate, which exceeds 50%, the outcome of metastatic and recurrent OS remains unchanged, with a survival rate <20%¹⁹. For spinal OS, the low incidence and proximity to pivotal structures make the cure challenging⁶. Thus, the need for developing novel therapies is urgent for OS treatment. Based on these findings, we focused on the underlying role of miR-520b in OS cells. The specific effects of miR-520b on cell proliferation,

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migration, and invasion were all explored. In addition, the possible signaling pathway involved in the modulation was further studied.

MATERIALS AND METHODS

OS Tissue Collection

Guardians of four patients (two males and two females, average age: 14.5 ± 1.3) with spinal OS agreed to join the study and signed the informed consent. OS tissues and adjacent nontumor tissues of OS patients were collected during the surgery, between 2014 and 2015, at The Affiliated Hospital of Qingdao University. The samples were quickly frozen in liquid nitrogen and stored at -80°C for further experiments. The process was approved by the ethics committee of The Affiliated Hospital of Qingdao University.

Cell Culture

Human fetal osteoblastic cell line (hFOB) and three kinds of OS cell lines, including Saos2, U2OS, and MG63, were all purchased from The Shanghai Institutes for Biological Sciences Cell Resource Center (Shanghai, P.R. China). hFOB cells were cultured in 1:1 mixture (Gibco, Gaithersburg, MD, USA) of Ham's F12 medium and Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (FBS; Invitrogen, Carlsbad, CA, USA) and 0.3 mg/ml G418 (Gibco). MG63, Saos2, and U2OS cells were cultured in DMEM supplemented with 10% FBS, except Saos2, whose final concentration of FBS was 15%. All the cells were maintained in a humidified atmosphere with 5% CO_2 at 37°C .

Cell Transfection

Lentivirus vectors carrying negative control miR (control), miR-520b precursor (miR-520b) or miR-520b inhibitor (anti-miR-520b), small interfering RNA targeting frizzled-8 (si-FZD8), and negative control of si-FZD8 (siNC) were all synthesized by GenePharma (Shanghai, P.R. China). Full-length cDNA of human FZD8 was subcloned into pc-DNA3.1 vector (Invitrogen), and the recombinant plasmid (pc-FZD8) was sequenced by Sangon (Shanghai, P.R. China). For the lentiviral infection, MG63 cells were seeded into a six-well plate and maintained for 24 h. The three recombinant lentivirus vectors were then infected into cells at a multiplicity of infection (MOI) of 10. The stable clones were selected using puromycin (Santa Cruz Biotechnology, Santa Cruz, CA, USA). Cell transfection of si-FZD8, siNC, or pc-FZD8 was performed in MG63 cells with Lipofectamine 2000 reagent (Invitrogen) in accordance with the supplier's instructions.

Cell Proliferation

Cell proliferation was evaluated by 3-(4, 5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide (MTT) and

colony formation assays. For the MTT assay, transfected MG63 cells (5×10^3 cells/well) were seeded into a 96-well plate and cultured in a humidified incubator with 5% CO_2 at 37°C for 4 days. At 1, 2, 3, and 4 days, 10 μl of MTT solution (5 mg/ml; Sigma-Aldrich, St. Louis, MO, USA) was added to each well, and the mixture was incubated for 4 h at 37°C . The medium was then removed, and 150 μl of dimethyl sulfoxide (DMSO) was added into each well. After oscillation and dissolution of crystals, the absorbance was recorded at 570 nm using a microplate reader (BioTek, Winooski, VT, USA). For the colony formation assay, MG63 cells (500 cells/well) were seeded into six-well plates and maintained for 48 h. The cells were then fixed by methanol for 20 min and stained with 0.5% crystal violet for 15 min. Subsequently, the number of colonies was counted under an inverted microscope (Olympus, Tokyo, Japan).

Cell Migration and Invasion Assay

Cell migration was assessed using a 24-well Transwell chemotaxis chamber (Costar, Corning, NY, USA) with a membrane (8- μm pore size). After transfection, 200 μl of cells in serum-free medium was filled into the upper chamber, while the lower chamber was filled with 600 μl of complete medium. A day later, cells that did not migrate were carefully removed from the upper membrane by a cotton swab. In the meantime, the cells that passed through the membrane were fixed and stained with hematoxylin. The migrated cells were then photographed and counted using an inverted microscope (Olympus). In terms of cell invasion, the process was the same as cell migration except that the membrane was precoated with 20 μl of Matrigel (BD Biosciences, San Jose, CA, USA). Each experiment was repeated three times.

Quantitative Reverse Transcription (qRT)-PCR

Total RNA was isolated with TRIzol reagent (Invitrogen) and DNase I (Promega, Madison, WI, USA) according to the manufacturer's instructions. The RNA was then reversely transcribed by MultiScribe reverse transcriptase and random hexamers (both from Applied Biosystems, Foster City, CA, USA) in line with the manufacturer's protocol. qRT-PCR was performed using the SYBR green mix (Applied Biosystems) in accordance with the supplier's protocol. Primers were designed and synthesized by Sangon. The relative expression of miR-520b and FZD8 was calculated using the $2^{-\Delta\Delta\text{Ct}}$ method²⁰. U6 acted as the housekeeping gene of miR-520b, and GAPDH acted as the housekeeping gene of FZD8.

Western Blot Analysis

The proteins were extracted using Nuclear and Cytoplasmic Protein Extraction Kit (Beyotime, Shanghai,

P.R. China) and phenylmethanesulfonyl fluoride (PMSF; Beyotime) according to the manufacturer's protocol. The concentration of proteins was then estimated by BCA™ Protein Assay Kit (Pierce, Appleton, WI, USA). Equivalent proteins were loaded onto 10% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) gels. Thereafter, the proteins were transferred to polyvinylidene fluoride (PVDF) membranes (Millipore, Billerica, MA, USA), followed by blocking with 5% skim milk (Nestlé, Shuangcheng, P.R. China). After rinsing, the membranes were incubated at 4°C overnight with primary antibodies against FZD8 (ab155093), cyclin D1 (ab137875), c-Myc (ab152146), β -catenin (ab6302), T-cell transcription factor 4 (TCF-4; ab185736), or GAPDH (ab128915) (all from Abcam, Cambridge, MA, USA). After rinsing again, the membranes were incubated with secondary antibody marked by horseradish peroxidase for 1 h at room temperature. The membranes were washed again and then transferred into the Bio-Rad ChemiDoc™ XRS system, followed by adding 200 μ l of Immobilon Western Chemiluminescent HRP Substrate (Millipore) to cover the membrane surface. The signals were captured using Image Lab™ Software (Bio-Rad, Hercules, CA, USA).

Statistical Analysis

All experiments were repeated three times. The results were presented as the mean \pm standard deviation (SD). Statistical analysis was performed using GraphPad Prism 5 software (GraphPad, San Diego, CA, USA). The p values were calculated using one-way analysis of variance (ANOVA) with Bonferroni's correction for comparison between three groups or two-tailed unpaired t -test for comparison between two groups. A value of $p < 0.05$ was defined as statistically significant.

RESULTS

miR-520b Is Downregulated in OS Tissues and Cell Lines

The difference in expression of miR-520b between tumor tissues and nontumor tissues in spinal OS as well as OS cells and normal cells was assessed by qRT-PCR. When compared to normal hFOB cells, miR-520b was significantly downregulated in Saos2 ($p < 0.01$), MG63 ($p < 0.001$), and U2OS ($p < 0.001$) cells (Fig. 1A). Similarly, miR-520b was significantly downregulated in spinal OS tissues compared with adjacent nontumor tissues ($p < 0.01$ or $p < 0.001$) (Fig. 1B). Therefore, we concluded that miR-520b was downregulated in OS.

Overexpression of miR-520b Suppresses Proliferation, Migration, and Invasion in OS Cells

To explore the effect of miR-520b on OS, we infected lentivirus vectors into MG63 cells to overexpress miR-520b. miR-520b was significantly overexpressed in the miR-520b group compared with the control group ($p < 0.001$) (Fig. 2A). Cell viability was markedly decreased by miR-520b overexpression compared with the control group at 2 ($p < 0.01$), 3 ($p < 0.001$), and 4 days ($p < 0.01$) after infection (Fig. 2B). Similarly, colony number, cell migration, and invasion were all decreased by miR-520b overexpression compared with the control group ($p < 0.01$) (Fig. 2C–E). These results suggested that miR-520b could inhibit cell proliferation, migration, and invasion.

Knockdown of miR-520b Promotes Proliferation and Migration in OS Cells

To validate the effect of miR-520b on OS cells, we also infected lentivirus vectors into MG63 cells to silence miR-520b. The expression of miR-520b was significantly

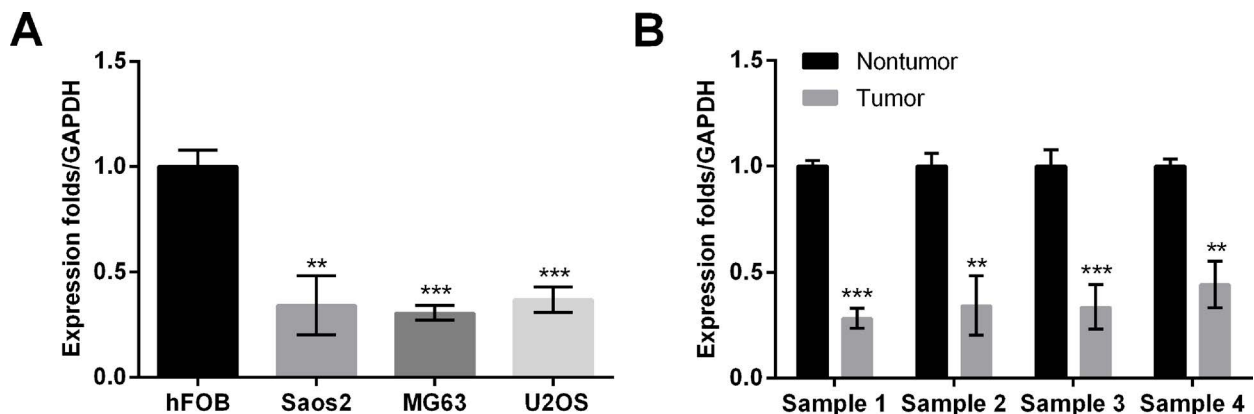


Figure 1. miR-520b is downregulated in OS tissues and cells. The expression of miR-520b was markedly reduced in OS tissues (A) and OS cells (B). Data are expressed as mean \pm standard deviation (SD) of at least three independent experiments. ** $p < 0.01$; *** $p < 0.001$. miR-520b, microRNA-520b; OS, osteosarcoma; hFOB, human fetal osteoblastic.

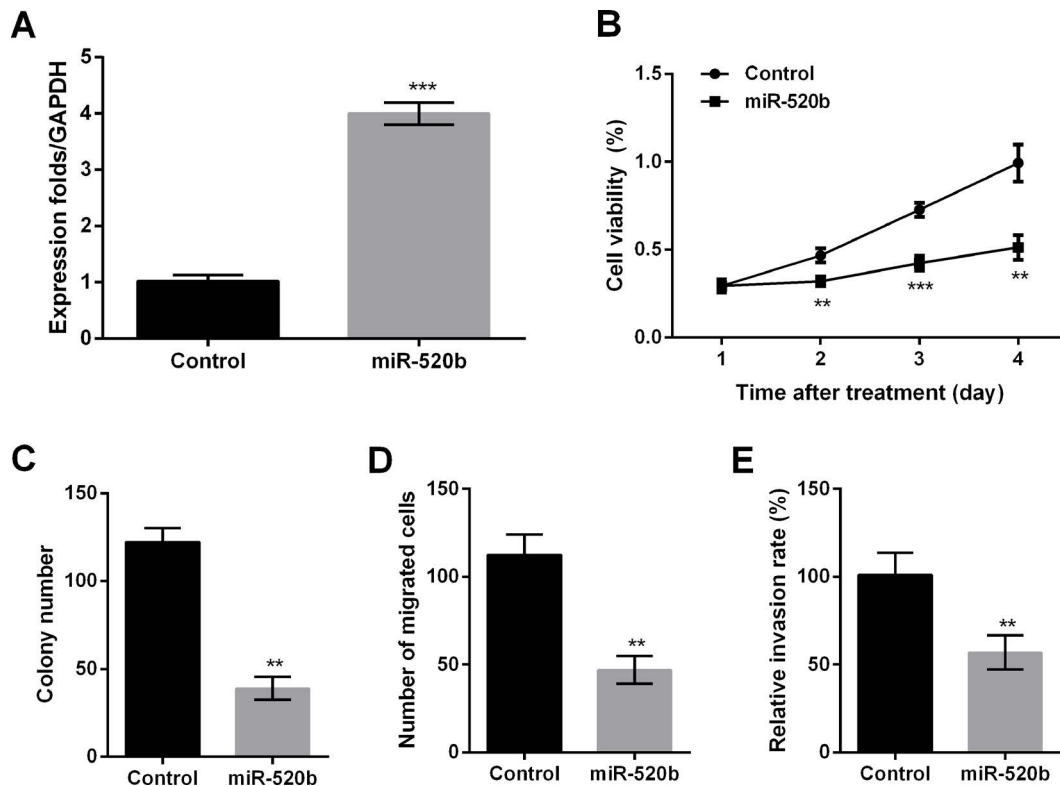


Figure 2. miR-520b inhibits cell proliferation, migration, and invasion in MG63 cells. (A) miR-520b levels by quantitative reverse transcription (qRT)-PCR. (B) Cell viability by MTT assay. (C) Colony number was reduced by miR-520b. Cell migration (D) and invasion (E) were both repressed by miR-520b. Data are expressed as mean \pm SD of at least three independent experiments. ** $p < 0.01$; *** $p < 0.001$.

reduced by infection of anti-miR-520b compared with the control group ($p < 0.001$), indicating that miR-520b was successfully silenced (Fig. 3A). Cell viability was markedly enhanced by miR-520 knockdown compared to the control group at 2 ($p < 0.05$), 3 ($p < 0.01$), and 4 days ($p < 0.05$) after infection (Fig. 3B). Likewise, the colony number and cell migration were both obviously increased by miR-520 knockdown when compared to the control group ($p < 0.001$) (Fig. 3C and D). These data proposed that miR-520b knockdown could promote cell proliferation and migration.

miR-520b Modulates Degradation of FZD8

To identify the influence of miR-520b on FZD8 expression, we infected different doses of miR-520b (10, 20, and 40 nM) and infected 40 nM miR-520b for different lengths of time (0, 4, 8, 12, and 24 h), followed by Western blot analysis of FZD8 expression. FZD8 expression was tapered off along with the concentration increase in miR-520b (Fig. 4A). FZD8 expression was also tapered off along with the increase in infection time (Fig. 4B). The miR-520b level was negatively correlated with FZD8 mRNA expression (Fig. 4C). Furthermore, the protein

expression of FZD8 in three OS cell lines and four clinical OS tissues were all obviously enhanced when compared with their respective controls, demonstrating just the opposite alteration compared with miR-520b levels (Fig. 4D and E). Thus, we drew the conclusion that miR-520b modulated degradation of FZD8.

miR-520b Inactivates the Wnt/ β -Catenin Signaling Pathway via FZD8

To reveal the underlying mechanism of miR-520b-associated regulations, the protein expression level of key proteins involved in the Wnt/ β -catenin signaling pathway was assessed by Western blot analysis. Cyclin D1, c-Myc, nuclear β -catenin, and TCF-4 were all obviously upregulated by miR-520b knockdown (Fig. 5A). These four proteins were all significantly downregulated by miR-520b overexpression (Fig. 5B). In addition, alteration of these four proteins induced by FZD8 silencing was the same as that induced by miR-520b overexpression (Fig. 5C). Simultaneously, cell viability was remarkably decreased by FZD8 silencing compared with the siNC group at 2 ($p < 0.01$), 3 ($p < 0.01$), and 4 days ($p < 0.05$) after transfection, indicating that the effect of FZD8 silencing on

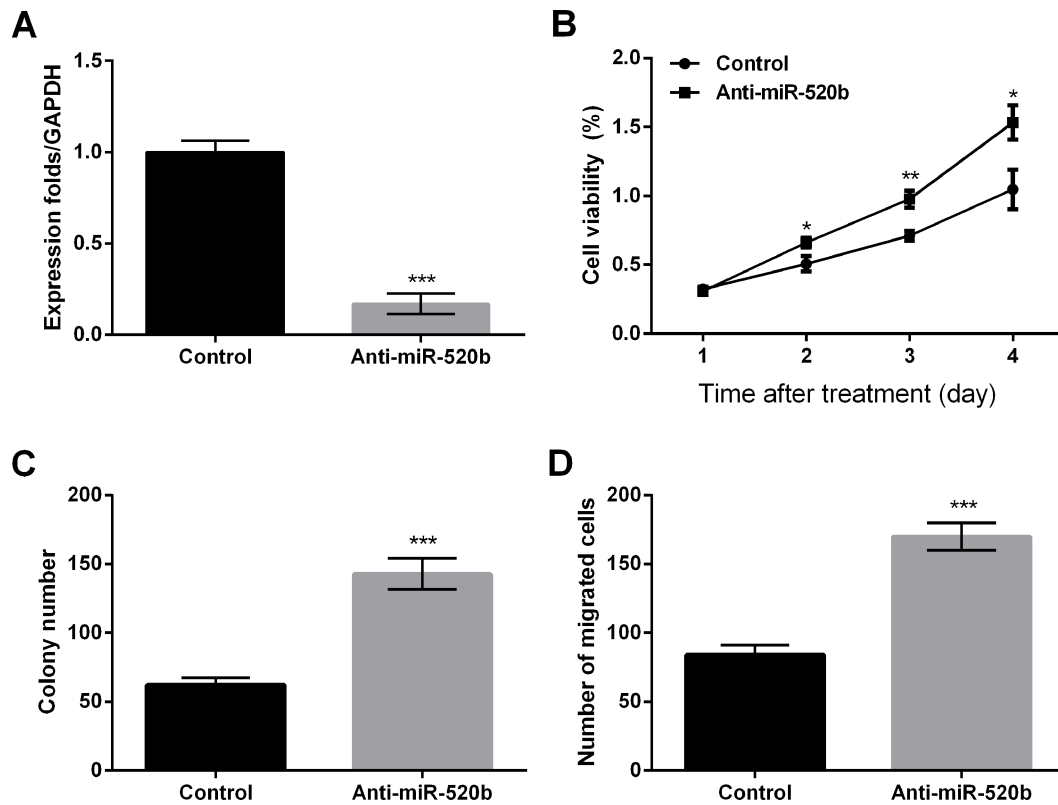


Figure 3. miR-520b silencing promotes cell proliferation and migration in MG63 cells. (A) miR-520b levels by qRT-PCR. (B) Cell viability by MTT assay. (C) Colony number was increased by miR-520b silencing. (D) Cell migration was promoted by miR-520b silencing. Data are expressed as mean \pm SD of at least three independent experiments. * p < 0.05; ** p < 0.01; *** p < 0.001. Anti-miR-520b, lentivirus vector carrying miR-520b inhibitor.

cell proliferation was the same as that of miR-520b overexpression. Furthermore, the effect of miR-520b overexpression on the expressions of cyclin D1, c-Myc, nuclear β -catenin, and TCF-4 was reversed by FZD8 overexpression (Fig. 5E). Taken together, we demonstrated that miR-520b inactivated the Wnt/ β -catenin signaling pathway through downregulation of FZD8.

DISCUSSION

Spinal OS represents 3.6%–14.5% of primary spinal tumors and 0.8%–3% of all OSs. Despite the development of modern medicine, the 5-year overall survival of spinal OS is just 30%–40%⁹. Therefore, exploration of the molecular mechanism involved in OS progression and a potential novel therapeutic target is very emergent. First, we interestingly identified that miR-520b was significantly downregulated in spinal OS tissues as well as in three OS cell lines. Thereafter, we found that miR-520b overexpression obviously inhibited cell proliferation, migration, and invasion, while the effect of miR-520b knockdown was just the opposite. Subsequently, miR-520b was revealed to modulate degradation of FZD8.

The final results showed that miR-520b inactivated the Wnt/ β -catenin signaling pathway through downregulation of FZD8.

miR-520b is reported to be involved in multiple types of cancer. Previous experiments performed using clinical tissues implied that miR-520b was downregulated in hepatocellular carcinoma¹⁵ and breast cancer tissues¹⁷. However, little research has been done to illustrate the expression level of miR-520b in spinal OS tissues and adjacent nontumor tissues. In our study, we first detected the miR-520b levels not only in spinal OS tissues but also in OS cell lines. Unsurprisingly, we found that the miR-520b levels were downregulated in both clinical spinal OS tissues and OS cell lines, which was consistent with the alterations in other types of cancer.

As miR-520b is reported to be a tumor suppressor, we further explored the role of miR-520b in cell proliferation, migration, and invasion of OS cells. After infection with different lentiviral vectors, we successfully obtained miR-520b-overexpressed or -silenced OS cells. The following experiments implied that miR-520b overexpression significantly inhibited cell proliferation, migration,

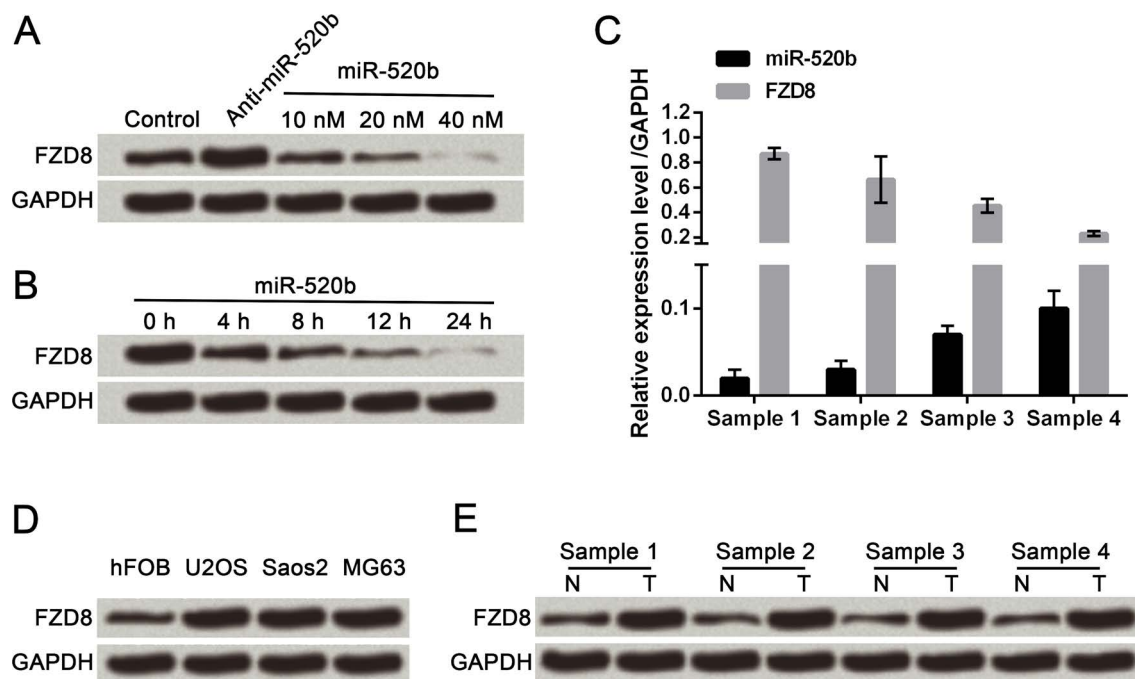


Figure 4. miR-520b modulates degradation of frizzled-8 (FZD8). (A) FZD8 was downregulated along with concentration increase in miR-520b. (B) FZD8 was downregulated along with time increase in miR-520b infection. (C) miR-520b negatively correlated with FZD8 mRNA expression. (D) FZD8 protein expression levels in hFOB, U2OS, Saos2, and MG63 cells. (E) FZD8 protein expression levels in spinal OS tissues and adjacent nontumor tissues. Data are expressed as mean \pm SD of at least three independent experiments.

and invasion. The influence of miR-520b knockdown was opposite to overexpression. The results of our study agreed with previous literature. Li et al. suggested that miR-520b represses proliferation and migration by targeting EGFR in gastric cancer¹⁵. Lu et al. reported that miR-520b inhibited the malignancy of head and neck cancer by targeting CD44 molecules²⁰.

According to previous investigations, modulation of miR-520b on cancer cells usually functions by regulation of another gene. Thus, we subsequently explored the possible correlation between miR-520b and other genes. A previous study reported that FZD8 acts as a putative therapeutic target in lung cancer, along with promotion of cell proliferation²¹. Furthermore, a secreted drug that consists of the ligand-binding domain of FZD8 has been proven to have an antitumor influence on animals²². Therefore, we infected different doses of miR-520b into OS cells for different infection times, followed by evaluation of FZD8 expression. The results demonstrated that the FZD8 expression was reduced in a dose- and time-dependent manner. Additionally, the data of RNA expression levels revealed that miR-520b was a negative control factor for FZD8. Comparison of the protein expression levels of FZD8 between normal cells and OS cells as well as spinal OS tissues and adjacent nontumor tissues suggested that FZD8 was highly expressed in

OS, which was opposite to that of miR-520b expression. Taken together, we hypothesized that miR-520b could modulate FZD8 degradation.

FZD8, one of the FZD family of receptors that act as Wnt protein receptors, is a crucial receptor for the Wnt/ β -catenin signaling pathway^{23,24}. In the canonical Wnt/ β -catenin signaling pathway, Wnt binds to the FZD receptor and activates the “destruction complex” of β -catenin, preventing phosphorylation and ubiquitination of β -catenin²⁵. As a consequence, β -catenin is accumulated and translocates to the nucleus where β -catenin complexes with TCF/leukemia enhancer factor 1 (LEF-1), leading to the transcription of downstream target genes, including cyclin D1 and c-Myc^{26,27}. Cyclin D1 is important for the G₁ to S transition and is involved in tumor progression and metastasis in OS²⁸. c-Myc is well studied and has been shown to regulate large numbers of genes involved in the cell proliferation of various types of cancer, including OS²⁹. In addition, suppression of the Wnt/ β -catenin signaling pathway is reported to inhibit cell migration and invasion of OS cells³⁰. Hence, we further explored the effect of miR-520b on the Wnt/ β -catenin signaling pathway through protein expression of cyclin D1, c-Myc, nuclear β -catenin, and TCF-4. Results demonstrated that the Wnt/ β -catenin pathway was activated by miR-520b silencing but inactivated by miR-520b overexpression.

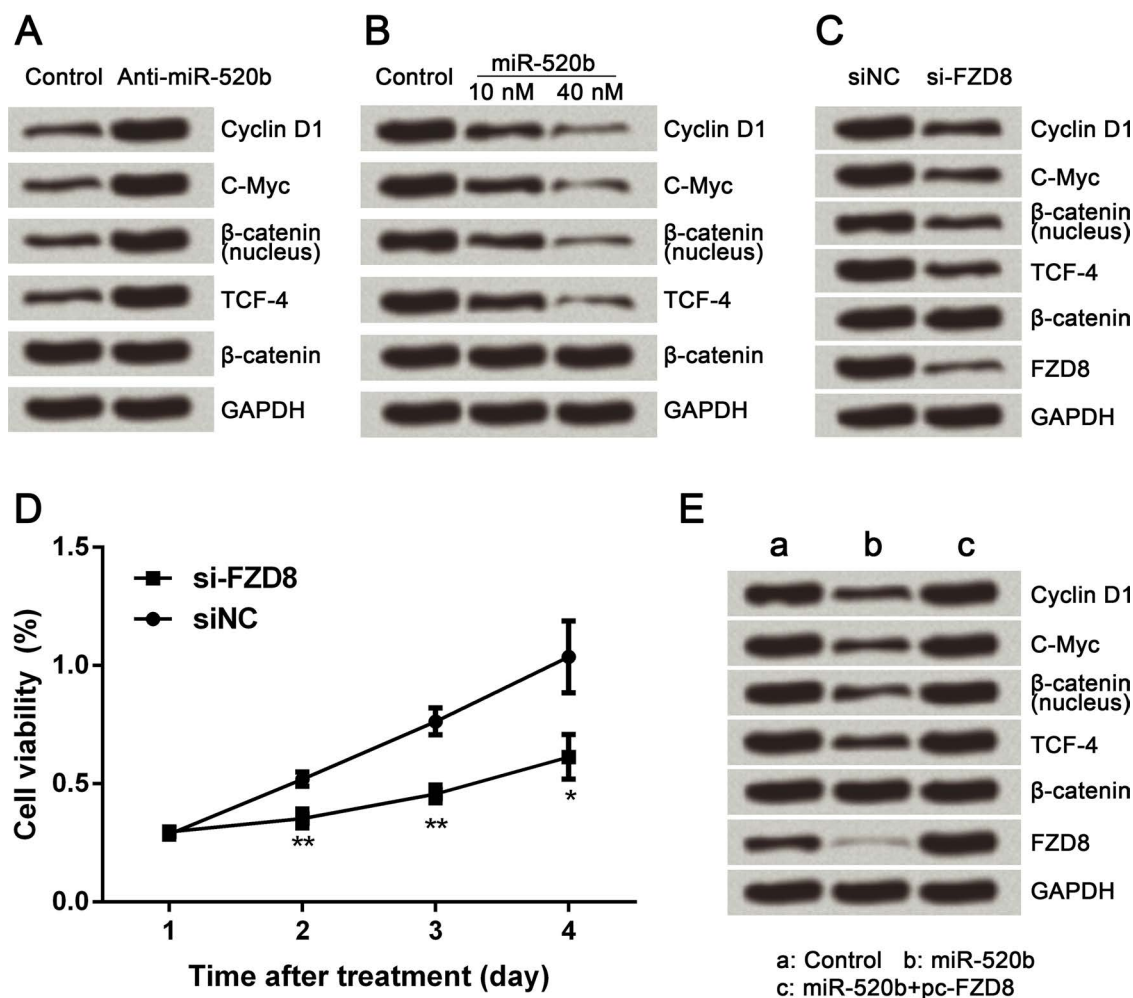


Figure 5. miR-520b inactivates the Wnt/ β -catenin signaling pathway through FZD8 downregulation. (A) Protein expression levels of cyclin D1, c-Myc, nuclear β -catenin, and TCF-4 were all enhanced by miR-520b inhibitor. (B) Protein expression levels of cyclin D1, c-Myc, nuclear β -catenin, and TCF-4 were all decreased by miR-520b overexpression. (C) Protein expression levels of cyclin D1, c-Myc, nuclear β -catenin, and TCF-4 were all decreased by FZD8 silencing. (D) Cell viability was decreased by FZD8 silencing. (E) Effect of miR-520b overexpression on protein expression levels of cyclin D1, c-Myc, nuclear β -catenin, and TCF-4 was reversed by FZD8 overexpression. Data are expressed as mean \pm SD of at least three independent experiments. * $p < 0.05$; ** $p < 0.01$. TCF4, T-cell transcription factor 4; si-FZD8, small interfering RNA targeting FZD8; siNC, negative control of si-FZD8; pc-FZD8, pcDNA3.1 vector carrying full-length cDNA of FZD8.

In addition, FZD8 knockdown inactivated the signaling pathway and inhibited cell proliferation. Meanwhile, FZD8 overexpression could abrogate the effect of miR-520b overexpression.

In conclusion, miR-520b was downregulated in spinal OS tissues and OS cell lines when compared with their respective controls. In OS cells, miR-520b could inhibit cell proliferation, migration, and invasion, while its knockdown showed the opposite results. Furthermore, miR-520b inactivated the Wnt/ β -catenin signaling pathway by downregulation of FZD8. The results provide novel clues for targeting therapy for OS. Deeper studies should be done to reveal the details between miR-520b and FZD8.

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